



**HAL**  
open science

# Epidemiological review of *Toxoplasma gondii* infection in humans and animals in Portugal

A.P. Lopes, J.P. Dubey, Marie-Laure Dardé, L. Cardoso

► **To cite this version:**

A.P. Lopes, J.P. Dubey, Marie-Laure Dardé, L. Cardoso. Epidemiological review of *Toxoplasma gondii* infection in humans and animals in Portugal. *Parasitology*, 2014, 141 (13), pp.1699-1708. 10.1017/S0031182014001413 . hal-01205307

**HAL Id: hal-01205307**

**<https://unilim.hal.science/hal-01205307>**

Submitted on 28 Aug 2020

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

# Epidemiological review of *Toxoplasma gondii* infection in humans and animals in Portugal

A. P. LOPES<sup>1,2\*</sup>, J. P. DUBEY<sup>3</sup>, M.-L. DARDÉ<sup>4,5,6</sup> and L. CARDOSO<sup>1,7</sup>

<sup>1</sup> Department of Veterinary Sciences, School of Agrarian and Veterinary Sciences, University of Trás-os-Montes e Alto Douro (UTAD), Quinta de Prados, 5000-801 Vila Real, Portugal

<sup>2</sup> Centro de Ciência Animal e Veterinária (CECAV), UTAD, Vila Real, Portugal

<sup>3</sup> United States Department of Agriculture, Agricultural Research Service, Beltsville Agricultural Research Center, Animal Parasitic Diseases Laboratory, Building 1001, BARC-East, Beltsville, Maryland 20705-2350, USA

<sup>4</sup> Centre National de Référence (CNR) Toxoplasmose/Toxoplasma Biological Resource Center (BRC), Limoges, France

<sup>5</sup> INSERM UMR 1094, Tropical Neuroepidemiology, Limoges, France

<sup>6</sup> University of Limoges, School of Medicine, Institute of Neuroepidemiology and Tropical Neurology, 2 avenue Martin Luther King, 87042 Limoges, France

<sup>7</sup> Parasite Disease Group, Instituto de Biologia Molecular e Celular (IBMC), Universidade do Porto, Oporto, Portugal

## SUMMARY

Toxoplasmosis is a worldwide zoonosis. However, data from Portugal are limited and a considerable part of the literature is in Portuguese. Currently, the rate of congenital infection in Portugal is unknown, and almost nothing is known of sequelae of congenital toxoplasmosis. There is no recent general population-based serological survey of *Toxoplasma gondii* in humans in Portugal. In addition, there is little information on genetic characteristics of *T. gondii* in animals and humans. In the present paper, we review prevalence, clinical spectrum and epidemiology of *T. gondii* in humans and animals in Portugal. This knowledge should be useful to biologists, public health workers, physicians and veterinarians.

Key words: *Toxoplasma gondii*, toxoplasmosis, humans, animals, epidemiology, Portugal.

## INTRODUCTION

The earliest paper we found on toxoplasmosis in Portugal is that of Campos (1951), which was written in Portuguese. Many other papers are also in Portuguese and some important data are buried in unpublished dissertations and theses, not available to a wider audience. Here we review the epidemiology, prevalence and clinical manifestations of *Toxoplasma gondii* infection in humans and animals from Portugal. Topics for future research are also suggested in order to encourage international collaboration.

## METHODS FOR PRESENT REVIEW

A review of the literature available until May 2014 was carried out by searching the PubMed database with reference combination of the following keywords: '(*Toxoplasma gondii* OR toxoplasmosis) AND Portugal'. Our initial search indicated 52 references, out of which 29 references on *T. gondii* infection or toxoplasmosis in humans and/or animals from Portugal were selected. Databases of universities, research institutes and scientific societies and the reference lists of all the retrieved articles were also

searched for published reports deemed as relevant to this review. After this, three additional papers were retrieved from non-indexed but peer-reviewed Portuguese journals. A set of 32 papers is reviewed in the main document. Four abstracts from the proceedings of national conferences, another abstract from the proceedings of an international conference, four master's dissertations and two doctoral theses also accounted for the scientific literature used to prepare the present review. That information is provided as a supplementary file (Supplement 1 – in Online version only).

Detailed historical, serological, parasitological, clinical and additional information on *T. gondii* infections in humans and other animals are summarized in tables (Tables 1–4) throughout the review. Fortunately, most studies used the commercially available modified agglutination test (MAT) to detect *T. gondii* antibodies in human and animal sera. This is the same test as described by Desmonts and Remington (1980) and Dubey and Desmonts (1987). It has been widely used to determine *T. gondii* seroprevalence in humans and animals (Dubey, 2010). This test assays only immunoglobulin (Ig) G antibodies, because the mercaptoethanol needed in the test destroys IgM and IgM-like antibodies. Cut-off values for MAT are listed wherever the authors provided the information. Data by other tests used is also provided.

\* Corresponding author. Departamento de Ciências Veterinárias Universidade de Trás-os-Montes e Alto Douro (UTAD), 5000-801 Vila Real, Portugal. E-mail: aplopes@utad.pt

Table 1. Seroprevalence of *T. gondii* infection in humans and domestic animals from Portugal

Host	Years	Region	Population	No. tested	Serological test (cut-off titre)	Prevalence (%)	Reference
Humans ( <i>Homo sapiens</i> )	1981	L	Pregnant women	686	MAT (20)	64.3	Antunes <i>et al.</i> (1981)
	1979–1980	N, C, At, L and Ag	General population	1675	IFAT (40)	47.0	Ángelo (1983)
Cat ( <i>Felis catus</i> )	2009–2010	N	Women of childbearing age	401	CMIA	24.4	Lopes <i>et al.</i> (2012)
	2003–2005	L	Stray	231	MAT (80)	24.2	Duarte <i>et al.</i> (2010)
	2004–2005	N	Domestic	204	MAT (20)	35.8	Lopes <i>et al.</i> (2008)
	2007–2008	L	Domestic	215	MAT (40)	20.5	Esteves <i>et al.</i> (2014)
	2009–2010	L	Stray	423	MAT (20)	44.2	Waap <i>et al.</i> (2012)
	2008–2009	N	Domestic	673	MAT (20)	38.0	Lopes <i>et al.</i> (2011a)
Dog ( <i>Canis familiaris</i> )	2008–2010	N	ND	161	MAT (100)	7.5	Lopes <i>et al.</i> (2013a)
Cattle ( <i>Bos taurus</i> )	2005–2006	N	Farmed	1467	MAT (20)	17.1	Sousa <i>et al.</i> (2009)
Sheep ( <i>Ovis aries</i> )	2008–2010	N	ND	119	MAT (20)	33.6	Lopes <i>et al.</i> (2013a)
	2008–2010	N	ND	184	MAT (20)	18.5	Lopes <i>et al.</i> (2013a)
Goat ( <i>Capra hircus</i> )	2004–2005	N	ND	333	MAT (20)	15.6	de Sousa <i>et al.</i> (2006)
Pig ( <i>Sus scrofa</i> )	2007–2008	C and L	Domestic	381	MAT (40)	7.1	Esteves <i>et al.</i> (2014)
Horse ( <i>Equus caballus</i> )	2008–2010	N	ND	254	MAT (20)	9.8	Lopes <i>et al.</i> (2013a)
	2008–2010	N	ND	173	MAT (20)	13.3	Lopes <i>et al.</i> (2013b)
Chicken ( <i>Gallus domesticus</i> )	Not stated	N and C	Free-range	225	MAT (10)	27.1	Dubey <i>et al.</i> (2006)

Ag, Algarve; At, Alentejo; C, Center; CMIA, chemiluminescence microparticle immunoassay; IFAT, indirect fluorescent antibody test; L, Lisbon; MAT, modified agglutination test; N, North; ND, not defined.

Table 2. Antibody titres of cats from Portugal tested by the MAT

No. of cats	Source	No. (%) of seropositive cats with MAT titres of:						Reference	
		20	40	60	80	160	180		≥ 320
194	Stray <sup>a</sup>				47 (24.2)				Duarte <i>et al.</i> (2010)
207	Pets <sup>b,c</sup>	3 (3.9)	18 (23.7)					55 (72.4)	Lopes <i>et al.</i> (2008)
215	Pets <sup>c</sup>		4 (1.9)				6 (2.8)	34 (15.8)	Esteves <i>et al.</i> (2014)
540	Stray <sup>d</sup>	24 (12.8)		5 (2.7)			14 (7.5)	144 (77.0)	Waap <i>et al.</i> (2012)

MAT, modified agglutination test.

<sup>a</sup> Catch and neuter programme in Lisbon.

<sup>b</sup> From individual owners.

<sup>c</sup> Veterinary clinics.

<sup>d</sup> Trapped by municipality.

Table 3. Seroprevalence of *T. gondii* infection in wild animals from Portugal sampled between 2008 and 2012

CLASS/order/species	Region	No. tested	% positive <sup>a</sup>	Reference
<i>BIRDS/Accipitriformes</i>				
Black kite ( <i>Milvus migrans</i> )	N	1	100	Lopes <i>et al.</i> (2011b)
Booted eagle ( <i>Hieraetus pennatus</i> )	N and C	2	50.0	Lopes <i>et al.</i> (2011b)
Common buzzard ( <i>Buteo buteo</i> )	N and C	26	69.2	Lopes <i>et al.</i> (2011b)
Eurasian sparrowhawk ( <i>Accipiter nisus</i> )	N	2	50.0	Lopes <i>et al.</i> (2011b)
Griffin vulture ( <i>Gyps fulvus</i> )	C	1	0.0	Lopes <i>et al.</i> (2011b)
Northern goshawk ( <i>Accipiter gentilis</i> )	N and C	3	100	Lopes <i>et al.</i> (2011b)
Short-toed eagle ( <i>Circaetus gallicus</i> )	C	2	0.0	Lopes <i>et al.</i> (2011b)
<i>BIRDS/Ciconiiformes</i>				
White stork ( <i>Ceconia ceconia</i> )	C	4	0.0	Lopes <i>et al.</i> (2011b)
<i>BIRDS/Columbiformes</i>				
Common pigeon ( <i>Columba livia</i> )	L	695	4.6	Waap <i>et al.</i> (2008)
	L	1507	2.6	Waap <i>et al.</i> (2012)
<i>BIRDS/Falconiformes</i>				
Peregrine falcon ( <i>Falco peregrinus</i> )	N	1	0.0	Lopes <i>et al.</i> (2011b)
<i>BIRDS/Strigiformes</i>				
Common barn owl ( <i>Tyto alba</i> )	N and C	2	0.0	Lopes <i>et al.</i> (2011b)
Eurasian eagle owl ( <i>Bubo bubo</i> )	N and C	3	33.3	Lopes <i>et al.</i> (2011b)
Tawny owl ( <i>Strix aluco</i> )	N	5	20.0	Lopes <i>et al.</i> (2011b)
<i>MAMMALS/Artiodactyla</i>				
European roe deer ( <i>Capreolus capreolus</i> )	N	1	0.0	Lopes <i>et al.</i> (2011b)
Wild boar ( <i>Sus scrofa</i> )	N	8	100	Lopes <i>et al.</i> (2011b)
	N	97	20.6	Coelho <i>et al.</i> (2014)
<i>MAMMALS/Carnivora</i>				
Common genet ( <i>Genetta genetta</i> )	N and C	2	100	Lopes <i>et al.</i> (2011b)
European badger ( <i>Meles meles</i> )	N and C	2	50.0	Lopes <i>et al.</i> (2011b)
Iberian wolf ( <i>Canis lupus signatus</i> )	N	1	100	Lopes <i>et al.</i> (2011b)
Red fox ( <i>Vulpes vulpes</i> )	N and C	6	100	Lopes <i>et al.</i> (2011b)

C: Center; N: North.

<sup>a</sup> Modified agglutination test (cut-off titre: 20).

Portugal, officially the Portuguese Republic, is located in Southwestern Europe. The country is located in the western part of the Iberian Peninsula, where it is bordered to the north and east by Spain and to the south and west by the Atlantic Ocean. The Portuguese territory has a total area of 92 090 km<sup>2</sup> and comprises a mainland and two insular autonomous regions: the archipelagos of Azores and Madeira, in the Northern Atlantic. Portugal has a population of around 10.8 million people, and joined the European Union in 1986. In the present work the country was divided according to the European

Union Nomenclature of Units for Territorial Statistics (NUTS) II, namely North, Centre, Alentejo, Lisbon, Algarve, Azores and Madeira regions (Fig. 1).

#### TOXOPLASMOSIS IN HUMANS

##### *Prevalence of T. gondii infection*

Published information is limited to three surveys; two of them are more than 20 years old (Table 1) and reported high seroprevalence. Antunes *et al.* (1981)

Table 4. Detection of *T. gondii* in tissues of animals from Portugal

Species	Source (location)	Sample	No. tested	% positive	Reference
Cat ( <i>Felis catus</i> )	Free range (L)	Brain and muscle	71	35.2 <sup>a</sup>	Waap <i>et al.</i> (2012)
	Domestic (L)	Feces	45	35.6 <sup>b</sup>	Esteves <i>et al.</i> (2014)
	Domestic (L)	Brain	15	6.7 <sup>b</sup>	Esteves <i>et al.</i> (2014)
Pig ( <i>Sus scrofa</i> )	Domestic (N)	Brain and/or heart	37	40.5 <sup>c</sup>	de Sousa <i>et al.</i> (2006)
	Domestic (C, L)	Brain and diaphragm	62	14.5 <sup>b</sup>	Esteves <i>et al.</i> (2014)
Chicken ( <i>Gallus domesticus</i> )	Free range (N, C)	Brain, leg muscle and heart	19	63.2 <sup>c</sup>	Dubey <i>et al.</i> (2006)
Pigeon ( <i>Columba livia</i> )	Free range (L)	Brain	23	52.2 <sup>b,c</sup>	Waap <i>et al.</i> (2008)
	Free range (L)	Brain	20	65.0 <sup>a</sup>	Waap <i>et al.</i> (2012)

C: Center; L: Lisbon; N: North.

<sup>a</sup> *In vitro* isolation.

<sup>b</sup> PCR.

<sup>c</sup> Isolation by bioassay.

found a 0.8% yearly incidence of infection in pregnant women from Lisbon and suggested an incidence of congenital infection of around 0.3% per pregnancy.

In the study by Ângelo (1983) (Table 1), seroprevalence varied from 19.8% in people aged 8 months to 5 years old to much higher levels in the 6–15 years, 16–30 years, 31–45 years and older population (39.1, 55.1, 57.1 and 57.8%, respectively). The reasons for these increasing results are most likely associated with a cumulative exposure with age. Analysing the results obtained by geographical districts, it was found that the mean values of seroprevalence of human infection were higher in the North than in the South of the country (Alentejo), ranging from 56.4% in Viana do Castelo (North) to 33% in Évora (Alentejo).

In another report by Ângelo (2003), seroprevalence in the population varied from 35% in the South to 70% in the North; for women of childbearing age, these values were between 30 and 60%. There is no national data concerning the prevalence of infection in pregnant women or in newborns. Nevertheless, Ângelo (2003) suggested the figure of 11 cases of congenital infection per 10000 live births, with an estimated number of 124 children congenitally infected each year.

We recently studied seroprevalence and associated risk factors of *T. gondii* in 401 women of childbearing age from the North of Portugal (Lopes *et al.* 2012a). Out of these, 343 women attending the gynaecology and obstetrics services of the public Hospital Centre of Trás-os-Montes e Alto Douro, in Vila Real, and 58 women attending private clinics participated in the study. Antibodies (IgG) to *T. gondii* were detected in 98 (24.4%) out of 401; two (2.0%) simultaneously had IgM. Almost 50% of the women studied by Lopes *et al.* (2012a) did not have any knowledge regarding toxoplasmosis. Risk factors for *T. gondii* infection in decreasing order were: soil-related activities without gloves (odds ratio [OR] = 8.4), consumption of unwashed raw vegetables or fruit (OR = 7.6) and eating smoked or cured



Fig. 1. Regions of Portugal according to the NUTS II. (adapted from Wikipedia).

processed pork products (OR = 2.5). Consumption of raw or undercooked meat was not identified as a risk factor. Exposure to oocysts rather than tissue cysts in meat was the most important risk factor associated with *T. gondii* infection in women in this study.

There is no legislation in Portugal that requires screening for toxoplasmic infection in pregnant women; however, in 2000, guidelines about the prevention of toxoplasmosis were made available by the General Directorate of Health (Ângelo, 2003).

### *Clinical toxoplasmosis*

*Congenital.* To our knowledge no confirmed clinical case has been published on congenital toxoplasmosis in Portugal. Campos (1951) reported severe encephalitis associated with a progressive blindness in a 3·5-year-old child. The child died 3 years later and *Toxoplasma*-like tachyzoites were found in the brain at autopsy. It is not certain if the child was congenitally or postnatally infected.

*Human immunodeficiency virus (HIV).* An HIV-seropositive patient with visceral leishmaniasis was further diagnosed with cerebral toxoplasmosis by the presence of IgG antibodies to *T. gondii* (dye and agglutination tests) and clinical and computer tomography (CT) scan features (Antunes *et al.* 1987). The patient's condition improved after pyrimethamine and sulphadiazine therapy for toxoplasmosis and antimony for leishmaniasis. In the frame of a European multicentre clinical research programme, Antunes (1991) reported 16 additional cases of toxoplasmic encephalitis in HIV patients.

A presumptive diagnosis of toxoplasmosis was made by Miranda *et al.* (1992) in 20 out of 95 HIV patients seen at the S. João Hospital, Oporto, between 1985 and 1991, based on symptoms (headache in 11, consciousness alterations in eight, vomiting in seven, fever in six, asthenia in five, and seizures in three), antibodies to *T. gondii*, CT, and favourable response to anti-*T. gondii* therapy (pyrimethamine and sulphadiazine in 16 patients and pyrimethamine with clindamycin in the remaining four). Diagnosis was confirmed in one patient by finding tachyzoites in brain biopsy. Two patients died after 3 to 4 weeks of therapy, but post-mortem diagnosis was not made. The serological study of *T. gondii* infection included detection of specific antibodies by enzyme-linked immunosorbent assay (ELISA).

In a retrospective study, Costa-Gomes *et al.* (2012) provisionally diagnosed toxoplasmosis based on CT or magnetic resonance studies or both in 11 out of 196 HIV patients. Nothing was said of serologic tests use, if any. Similarly, in a paper evaluating the surgical techniques of obtaining biopsy of brainstem for diagnosis of various ailments, Gonçalves-Ferreira *et al.* (2003) mentioned two male HIV patients with focal infiltration in brainstem biopsied at a hospital in Lisbon: a 37-year old and a 45-year old (who also had concurrent *Cytomegalovirus* infection). No other details were given.

Although CT and serological examination are useful, definitive diagnosis of toxoplasmosis can only be made post-mortem or by biopsy examination. Other conditions, including lymphomas, can mimic toxoplasmosis, and the determination of Ig isotype and the magnitude of titre to *T. gondii* are not helpful in the differential diagnosis. In most HIV-infected patients clinical toxoplasmosis is a reactivation of

chronic infection, and most of these patients have antibodies to *T. gondii*. Favourable response to anti-toxoplasmic therapy is helpful in ante-mortem diagnosis of toxoplasmosis.

*Ocular.* The only report on ocular toxoplasmosis we found is that of Palmares *et al.* (1990). They reviewed case histories of 450 patients seen in a referral hospital in Northern Portugal. Toxoplasmosis-associated uveitis was diagnosed in 41 (9·1%) out of 450 patients seen from 1985 to 1989. The diagnosis criteria were the morphologic picture and seropositivity in any of the serological tests. The report is brief and there is no information regarding the serological tests used and the titres. Our concept of toxoplasmosis has changed during the last 25 years, when this information was presented at a symposium. For example, postnatally acquired toxoplasmosis is now increasingly recognized as cause of uveitis; previously most of it was considered due to congenital infection.

*Organ transplant patients.* Abecassis (1991) reported two cases of encephalitis by *T. gondii* among 60 patients who received bone marrow transplantations, between May 1987 and May 1991. The first case was a 24-year-old female patient bone marrow transplanted for severe aplastic anaemia, which had been diagnosed 4 years before. Six months after transplantation, fever with poor general condition, severe muscle atrophy and a convulsive episode arised. An axial CT scan revealed multiple ring lesions. The severe thrombocytopenia presented by the patient did not allow brain biopsy and the patient was treated with pyrimethamine and sulphadiazine without favourable evolution of lesions on CT. The patient died 3 weeks later; autopsy was not performed. To us, the diagnosis is at best suspicious. In the second case the patient was a 41-year-old woman, transplanted for chronic myeloid leukaemia in chronic phase, diagnosed 8 months before. Prior to bone marrow transplantation an evaluation had been done with both the recipient and the donor being seronegative for *T. gondii* infection by ELISA. However, there was a subsequent seroconversion of the recipient with the detection of IgG and IgM antibodies, reflecting a recent infection. Therapy with spiramycin was instituted. Approximately 3 weeks after bone marrow transplantation the patient developed unusual cardiac complications and fever. Cranial CT revealed extensive left ventricular intracerebral ring shape lesions. Given the patient's condition, a severe thrombocytopenia and the previous evidence of recent infection with *T. gondii*, therapy with sulphadiazine and pyrimethamine was initiated. A favourable development was observed with complete recovery without neurological sequels.

Da Cunha *et al.* (1993a, b) reported evidence of cerebral toxoplasmosis in a 42-year-old male patient

after a successful renal transplantation in Portugal. The patient had headache and loss of consciousness; CT scan revealed focal lesion in brain. The patient had persistent serum antibodies to *T. gondii* and anti-*Toxoplasma* therapy (clindamycin and pyrimethamine) was initiated. When clinical and imaging improvement was observed, secondary prophylaxis was initiated with dapson and pyrimethamine, with favourable evolution (da Cunha *et al.* 1993a, b).

## TOXOPLASMOSIS IN ANIMALS

### *Clinical*

Virtually nothing is known of clinical toxoplasmosis in animals in Portugal.

### *Serological and parasitological prevalence*

Data are summarized in Tables 1–4.

*Cat.* Seroprevalence in cats deserves special attention because of the epidemiological importance of cats. Of the four serological surveys in cats, three were from Lisbon (Tables 1 and 2). The fourth survey (Lopes *et al.* 2008) was in domestic cats from Trás-os-Montes e Alto Douro (Northeastern Portugal); 161 cats were from Vila Real, and the remaining 46 cats were from other contiguous municipalities (Table 1). However, data provided were not further distinguished with respect to localities. All four studies with cats used the same MAT test, the cut-off titres (20–80) were different, and therefore data are not strictly comparable (Tables 1 and 2).

Serological prevalence varied from 20 to 44%; the highest being from stray cats from Lisbon. The stray cats were captured from different areas of Lisbon by the municipal services. It is noteworthy that a very high percentage of cats from Portugal had high titres (Table 2). In the study by Waap *et al.* (2012), 68 (36.3%) out of 187 seropositive cats had MAT titres of 18000 or higher reaching as much as 162000. One of us has never encountered such high MAT titres from cats from other parts of the world (J. P. Dubey, personal observations). This result may be due to the *T. gondii* strains or very high environmental contamination leading to repeated infections in cats.

Risk factors such as age, gender, diet and lifestyle were assessed in these four surveys. Gender did not affect the seropositivity. Age was one of the main risk factors; seroprevalence increased with age. Age-related data were: 14.6% seropositivity in 2–11 months old *vs* 51.7% in 3- to 6-year-old cats (Lopes *et al.* 2008); 20.6% in cats younger than 1 year *vs* 56.9% in cats older than 5 years (Waap *et al.* 2012); 7.7% in cats younger than 2 years *vs* 36.5% in cats older than 3 years (Esteves *et al.* 2014). These data indicate post-natal exposure of cats to *T. gondii*. There is no information on congenital transmission

of *T. gondii* in cats in Portugal. Cats that were fed on diets including raw or undercooked viscera and/or meat also had a higher percentage (53.5%) of infection in comparison with animals receiving only commercial canned or dried food (22.9%; Lopes *et al.* 2008). Feline contact was not shown to be a statistically significant risk factor for seropositivity in logistic regression analysis (Lopes *et al.* 2008).

As of yet, viable *T. gondii* oocysts have not been demonstrated in cat feces in Portugal. Oocysts were not found by microscopic examination of feces of 231 cats from Lisbon (Duarte *et al.* 2010). Ferreira *et al.* (2011) detected *T. gondii*-like oocysts in one out of 20 cats from private homes from region of Alentejo, but without bioassay a definitive diagnosis could not be made. Esteves *et al.* (2014) reported the presence of *T. gondii* DNA in feces of 16 (35.5%) out of 45 cat stools from Lisbon. This is a very high rate of DNA positivity, but bioassay is necessary for a definitive diagnosis of *T. gondii* in feces. The demonstration of DNA should not be equated with the presence of viable *T. gondii* oocysts. *T. gondii* DNA was also detected in six (13.3%) out of 15 brains of cats.

Viable *T. gondii* was isolated from a relatively high number of seropositive cats (Table 4). Waap *et al.* (2012) homogenized 1 g of brain from each of 56 seropositive cats, and isolated *T. gondii* from 15 of them in cell cultures seeded with this undigested brain homogenate. From another set of 15 cats they compared isolation of *T. gondii* in cell culture from brains (undigested 1 g) and pepsin digest of heart, and limb muscle (amount used not specified); *T. gondii* was isolated from muscles of 10 and brains of six cats.

This high rate of isolation (i.e. 15 [26.8%] out of 56) using 1 g of brain indicates high density of parasites in the brains of apparently healthy cats in Lisbon. These results are of interest because the success of isolation of *T. gondii* in cell cultures seeded by tissues of asymptomatic animals is generally low (Dubey, 2010).

### *Food animals as sources of infection*

Poultry, pork, beef and mutton are the most important source of meat consumed by humans in Portugal. Little is known of the relative risk of *T. gondii* infection with respect to meat sold in grocery stores. Although *T. gondii* antibodies were found in 7.5% (Table 1) and 29.2% of cattle (Supplement 1, Table S1 – in Online version only), viable *T. gondii* has rarely been isolated from beef in other countries (Dubey, 2010). Thus, the role of beef in the epidemiology of *T. gondii* is uncertain.

In addition to data in Table 4, viable *T. gondii* was isolated from a bovine foetus (Canada *et al.* 2002). During the course of an investigation on *Neospora caninum* (a protozoan similar to *T. gondii*) bovine fetal tissues were bioassayed in mice. For this,

homogenates of fetal brains were inoculated intraperitoneally into mice and protozoa harvested from the peritoneal cavity of infected mice were seeded on to cell cultures. One of these isolates proved to be *T. gondii*. The infected fetus was 5-month gestational age from a Holstein cow from the Azores (Canada *et al.* 2002). Confirmation of *T. gondii* as the causative agent of the abortion was not performed since the brains were not examined histologically.

Among the three surveys in pigs, seroprevalence varied from 7.1 to 15.6% (Table 1). Age and management were the two main factors concerning prevalence. Seroprevalence in feeder pigs (market age pigs) is important with respect to public health, because these pigs for human consumption are generally 6–10 months old at the time of slaughter. In this age group 5% out of 180 pigs from North of Portugal had antibodies to *T. gondii*; these pigs were from 14 farms (Lopes *et al.* 2013a). Similar results (i.e. 18 [6.8%] out of 264) were obtained from pigs from a slaughterhouse in Lisbon (Esteves *et al.* 2014). Seroprevalence were higher in older pigs, suggesting post-natal infection. As expected, pigs raised under extensive management had higher rate of *T. gondii* infection (Lopes *et al.* 2013a; Esteves *et al.* 2014).

No information was available concerning transplacental transmission of *T. gondii*, because only a few neonatal pigs were tested in both of these surveys. Transplacental transmission of *T. gondii* in pigs is uncommon (Dubey, 2010). De Sousa *et al.* (2006) found seropositivity in 63 (15.6%) out of 333 pigs from the municipality of Vinhais, in Northeastern Portugal; these pigs were 1–4 years old and mainly used for traditional sausage production. Brains/hearts (50 g) of 37 of these seropositive pigs were bioassayed in mice (Table 4). Viable *T. gondii* strains (TgPiPr1-15) were isolated from 15 (40.5%) of these pigs (de Sousa *et al.* 2006). *Toxoplasma gondii* DNA was detected in 9 (14.5%: 7 [35.0%] out of 20 seropositive; 2 [4.8%] out of 42 seronegative) out of 62 brains and diaphragms of pigs sampled at a slaughterhouse in Lisbon (Esteves *et al.* 2014).

In addition to meat from domestic pigs, meat from wild boars may also be an important source of *T. gondii* infection in humans. Coelho *et al.* (2014) found *T. gondii* antibodies in 20 (20.6%) out of 97 wild boars from the North of Portugal. In Portugal there is some tradition of wild boar meat consumption. According to the official data, it is estimated that around 15 400 wild boars are hunted per year in Portugal. Additionally, this meat is widely used to make uncontrolled homemade sausages, prepared without any heat processing and, therefore, consumed raw, increasing the likelihood of *T. gondii* as a foodborne pathogen for humans (Coelho *et al.* 2014). Additionally, infected viscera and carcasses left by hunters may pose a threat to scavenging susceptible animals.

Nothing is known of the prevalence of *T. gondii* in intensively raised chickens. Seroprevalence of *T. gondii* in free range chickens (small farms) is more indicative of soil contamination, because birds feed from the ground, than as a food source for the main population. Seroprevalence in 225 free range chickens from 18 farms was 20.8% using a MAT titre of 20 (Dubey *et al.* 2006). Tissues (brain, leg muscle and heart) of 19 seropositive chickens were bioassayed in mice and viable *T. gondii* strains (TgCkPr1-16) were isolated from 16 of them (Table 4). Using the same MAT titre, seroprevalence was much lower (5.2%) in 96 field-bred chickens from different parts of Portugal (Supplement 1, Table S1 – in Online version only).

The ingestion of undercooked mutton may be a risk factor for *T. gondii* infection in humans, due to the high prevalence of *T. gondii* in sheep and goats in Portugal (Table 1). Sousa *et al.* (2009) found a 17.1% seroprevalence (Table 1) of *T. gondii* infection in 1467 sheep from 160 farms in Vinhais. They also determined that a MAT cut-off of 20 was optimal for serodiagnosis of *T. gondii* infection in sheep; 86.2% sensitivity, 89.1% specificity, and negative and positive predictive values of 90.3 and 84.5%, respectively were obtained at this titre. Age of 7 months or older was considered a risk factor for *T. gondii* infection in sheep studied by Lopes *et al.* (2013a). We reported 50% or higher seropositivity in sheep older than 6 months, but only 34 sheep were included in this age group.

#### *Contamination of environment with T. gondii*

There are no specific data on the contamination of the soil and environment with *T. gondii* oocysts in Portugal. However, the high seroprevalence of *T. gondii* in cats (Tables 1 and 2) suggests that environment is likely to be contaminated, because cats that are seropositive have shed oocysts (Dubey, 2010). Although *T. gondii* oocysts are shed for only 1–2 weeks in the life of the cat, millions of oocysts can be shed and they can survive outdoors for months.

Pigeons were used as sentinels for *T. gondii* oocyst contamination in the city of Lisbon, because pigeons feed from the ground and they are herbivores (Waap *et al.* 2008, 2012). Seroprevalence among pigeons from 33 sites ranged from 4.6 to 65%, suggesting a considerable contamination of several urban areas of the city with *T. gondii* oocysts. Nineteen out of these 33 sites were coincident with feeding sites from the study in 2008 (Waap *et al.* 2008). Infection levels within the flocks ranged between 4.8 and 21.1%. The proportion of positive flocks rose from 36.8% (7/19) to 42.1% (8/19) at the same feeding sites regarding the previous study (Waap *et al.* 2008). The proportion of seropositive pigeons varied depending on the trimester of the year when blood samples were collected, being higher in the first trimester (January–March)



with 4.7% of seropositivity in pigeon flocks. Isolation of *T. gondii* from pigeons in this study (Table 4) using brain homogenates proved to be very efficient, with an overall isolation rate of 65% (13/20) and an isolation success of 100% (12/12) when considering pigeons with titres  $\geq 180$ . This efficiency of isolation is noteworthy, because only 0.5 g of brain was used for isolation and cell cultures were seeded with undigested brain suspension.

The high seroprevalence of *T. gondii* in domestic and wild herbivores (Tables 1 and 3) also indicates that rural environment is also contaminated with oocysts. In 173 horses from the North of Portugal (Table 1), no statistical differences were found among the equine categories of gender (female, male and gelding), age (1.5–6, 7–12 and 13–30 years), type of housing (indoors and mixed/outdoors), ability (recreation, farming and sports) and clinical status (apparently healthy and sick) for both agents. These results indicate that horses in the North of Portugal are exposed to *T. gondii* and their tissues might harbour cysts. Although voluntary consumption of horse meat by people is not a common habit in Portugal, the risk of *T. gondii* transmission from horses to humans should not be neglected due to the recent contamination of beef with horse meat noticed in Europe (Lopes *et al.* 2013b).

Dogs have been used as sentinel animals for estimating *T. gondii* infection in the environment. Dogs are known to eat cat feces and roll over in cat feces. In the only report in dogs from Portugal, risk factors for *T. gondii* infection were age above 12 months (OR = 4.0), chance of eating birds or small mammals (OR = 4.0), housing exclusively outdoors (OR = 1.5), and diet including home-cooked food (OR = 3.0) or raw meat and viscera (OR = 7.7). Contact with other domestic animals was not identified as a risk factor. The major risk factor for *T. gondii* infection in dogs was the inclusion of raw meat or viscera in the diet (Lopes *et al.* 2011a).

#### *Genetic characterization of T. gondii strains from Portugal*

Only a small percentage of exposed adult humans or other animals develop clinical signs of disease. The severity of toxoplasmosis in immunocompetent hosts is a mix of host and parasitic factors (host variability, inoculum doses and parasite strain, among others). Recently, attention has been focused on the genetic variability among *T. gondii* isolates from apparently healthy and sick hosts. Severe cases of toxoplasmosis that have been reported in immunocompetent patients are considered to be due to infection with atypical *T. gondii* genotypes (Robert-Gangneux and Dardé, 2012). There is little information on genetic typing of *T. gondii* isolates circulating in Portugal. We are aware of only one characterization report of *T. gondii* from humans in Portugal (Supplement 1 –

in Online version only). An indirect indication of genotypes circulating in humans is provided through serotyping methods (Sousa *et al.* 2008). A serotyping based on detection of antibodies against GRA6 peptide found a majority of GRA6 Type II profile as in other European countries, but also a significantly higher prevalence of serotype I/III among Portuguese patients (15%) than among French patients (2%). These results might suggest a different epidemiological pattern of strains circulating in these two countries.

Information on genetic typing is summarized here. The quality of DNA is important for genetic typing and complete data can be obtained only from DNA extracted from large numbers of viable parasites, usually cell or mouse-cultured organisms. More limited information can be obtained on DNA extracted directly from tissues of asymptomatic animals. Humans become infected with *T. gondii* mostly by consumption of uncooked meat or the oocysts. Therefore, information on genotypes of isolates from animals, especially cats, is relevant to human infections.

Different methods have been used to type the isolates. Earlier information was obtained using polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) with only SAG2 marker; out of the 12 analysed isolates from chickens, eight were Type II and four were Type III (Dubey *et al.* 2006). Similar results were obtained with isolates from pigs. Using SAG2 and five microsatellite markers, 11 out of the 15 isolates from pigs were Type II and four isolates were Type III. These results were confirmed by using serotyping using the GRA6 and GRA7 derived peptides (Sousa *et al.* 2010). Waap *et al.* (2008) characterized *T. gondii* DNA obtained directly from brain tissue from 12 out of 23 seropositive pigeons by five microsatellites; nine strains were Type II, two strains were Type III and one Type I.

Type II is the most prevalent *T. gondii* in Europe and North America (Su *et al.* 2012). Microsatellite and PCR-RFLP typing of the B1 strain from aborted bovine foetus (Canada *et al.* 2002) revealed a Type I strain, which is an unusual occurrence in the North of Portugal (A. P. Lopes, personal observations). This genotype is very rarely isolated in Europe, and its isolation on the Azores, may reflect a possible importation from the American continent.

#### *Concluding remarks and future directions*

Since congenital toxoplasmosis is of great concern to public health, with important medical, social and even economic consequences, and since infection acquired postnatally is seldom accompanied by clinical manifestations, knowledge on the epidemiology of infection in humans is of particular significance in the battle against this parasite. The

knowledge of the main sources of infection for the human and animal populations in Portugal may allow the development of more effective measures for primary prevention of infection in the human groups at higher risk, including pregnant women, women in childbearing age and HIV patients, thus minimizing congenital and cerebral toxoplasmosis. There is a need for a population-based survey of *T. gondii* in humans in different regions and persons of different ethnicity.

Among other topics of interest, it would be important to deepen the study in wild carnivores, such as the European wild cat (*Felis silvestris*) and the Iberian lynx (*Linx pardinus*), in order to identify possible sources of infection that can take part in the food chain of these predators, such as birds and wild rabbits, and to verify the impact of this protozoal infection on the decline in the population of those endangered species. Furthermore, as Portugal produces livestock meat for internal consumption and for export, isolation of *T. gondii* from meat and further characterization of the isolates will be needed to better understand the risk that especially sheep infection may represent for the human health.

The information summarized here should be of help to veterinarians, physicians, public health workers and biologists.

#### SUPPLEMENTARY MATERIAL

To view supplementary material for this article, please visit <http://dx.doi.org/10.1017/S0031182014001413>.

#### ACKNOWLEDGEMENTS

The findings and conclusions in this report are those of the authors and do not necessarily represent the views of the U.S. Department of Agriculture.

#### FINANCIAL SUPPORT

This work was sponsored by the Foundation for Science and Technology (FCT), Ministry of Education and Science, Portugal, under the Project PEst-OE/AGR/UI0772/2014.

#### REFERENCES

**Abecassis, M. M.** (1991). Complicações pouco habituais da transplantação de medula óssea (TMO). Experiência da Unidade de TMO do Instituto Português de Oncologia de Francisco Gentil, Centro de Lisboa. *Acta Médica Portuguesa* 4(Suppl. 1), S37–S38.

**Ângelo, M. H.** (1983). Prevalência dos anticorpos antitoxoplasmose. *Arquivos do Instituto Nacional de Saúde* 8, 105–111.

**Ângelo, M. H.** (2003). Dispositions légales et stratégies préventives de la toxoplasmose congénitale au Portugal. *Archives de Pédiatrie* 10, 25–26.

**Antunes, F.** (1991). Perspectivas terapêuticas da infecção pelo VIH e das infecções oportunistas. Projectos de investigação clínica multicêntricos na CEE. *Acta Médica Portuguesa* 4(Suppl. 1), 67–68.

**Antunes, F., Bacelar, F., Janz, J. G., Saraiva, J. and Araújo, F. C.** (1981). Prevalência e incidência da toxoplasmose adquirida em 686 grávidas

da região de Lisboa: sua influência na incidência da toxoplasmose congénita. *O Médico* 101, 904–912.

**Antunes, F., Carvalho, S., Tavares, L., Botas, J., Fortes, M., del Rio, A. M., Dutschmann, L., Costa, A., Abranches, P., Pereira, C. S., Paiva, J. E. D., Araújo, F. C. and Baptista, A.** (1987). Visceral leishmaniasis recrudescence in a patient with AIDS. *Transactions of the Royal Society of Tropical Medicine and Hygiene* 81, 595.

**Campos, J. O.** (1951). Toxoplasmose: nota sobre o primeiro caso observado em Portugal. *Revista Clínica do Instituto Maternal de Lisboa* 3, 9–23.

**Canada, N., Meireles, C. S., Rocha, A., da Costa, J. M., Erickson, M. W. and Dubey, J. P.** (2002). Isolation of viable *Toxoplasma gondii* from naturally infected aborted bovine fetuses. *Journal of Parasitology* 88, 1247–1248.

**Coelho, C., Vieira-Pinto, M., Faria, A. S., Vale-Goncalves, H., Veloso, O., Paiva-Cardoso, M. N., Mesquita, J. R. and Lopes, A. P.** (2014). Serological evidence of *Toxoplasma gondii* in hunted wild boar from Portugal. *Veterinary Parasitology* 202, 310–312.

**Costa-Gomes, B., Nunes, J., Pinto, J., Gouveia, P. and Pais, R. P.** (2012). Neuroimagem na infecção Vírus da Imunodeficiência Humana. *Acta Médica Portuguesa* 25(Suppl. 1), S7–S12.

**da Cunha, S., Ferreira, E., Ramos, I., Martins, R., de Freitas, L., Borges, J. L., Côrte-Real, R., Mota, A., Meliço-Silvestre, A. and Furtado, A. L.** (1993a). Cerebral toxoplasmosis after renal transplantation. Case report and review. *Acta Médica Portuguesa* 7(Suppl. 1), S61–S66.

**da Cunha, S., Ferreira, E., Ramos, I., Martins, R., de Freitas, L., Borges, J. L., Côrte-Real, R., Mota, A., Meliço-Silvestre, A. and Linhares-Furtado, A.** (1993b). Toxoplasmose cerebral em transplantado renal. Caso clínico e revisão da literatura. *Acta Médica Portuguesa* 6, 157–163.

**Desmonts, G. and Remington, J. S.** (1980). Direct agglutination test for diagnosis of *Toxoplasma* infection: method for increasing sensitivity and specificity. *Journal of Clinical Microbiology* 11, 562–568.

**de Sousa, S., Ajzenberg, D., Canada, N., Freire, L., da Costa, J. M., Dardé, M. L., Thulliez, P. and Dubey, J. P.** (2006). Biologic and molecular characterization of *Toxoplasma gondii* isolates from pigs from Portugal. *Veterinary Parasitology* 135, 133–136.

**Duarte, A., Castro, I., Pereira da Fonseca, I. M., Almeida, V., Madeira de Carvalho, L. M., Meireles, J., Fazendeiro, M. I., Tavares, L. and Vaz, Y.** (2010). Survey of infectious and parasitic diseases in stray cats at the Lisbon Metropolitan Area, Portugal. *Journal of Feline Medicine and Surgery* 12, 441–446.

**Dubey, J. P.** (2010). *Toxoplasmosis of Animals and Humans*, 2nd Edn. CRC Press, Boca Raton, FL, USA.

**Dubey, J. P. and Desmonts, G.** (1987). Serological responses of equids fed *Toxoplasma gondii* oocysts. *Equine Veterinary Journal* 19, 337–339.

**Dubey, J. P., Vianna, M. C., Sousa, S., Canada, N., Meireles, S., Correia da Costa, J. M., Marcet, P. L., Lehmann, T., Dardé, M. L. and Thulliez, P.** (2006). Characterization of *Toxoplasma gondii* isolates in free-range chickens from Portugal. *Journal of Parasitology* 92, 184–186.

**Esteves, F., Aguiar, D., Rosado, J., Costa, M. L., de Sousa, B., Antunes, F. and Matos, O.** (2014). *Toxoplasma gondii* prevalence in cats from Lisbon and in pigs from centre and south of Portugal. *Veterinary Parasitology* 200, 8–12.

**Ferreira, F. S., Pereira-Baltasar, P., Parreira, R., Padre, L., Vilhena, M., Távora Tavira, L., Atouguia, J. and Centeno-Lima, S.** (2011). Intestinal parasites in dogs and cats from the district of Évora, Portugal. *Veterinary Parasitology* 179, 242–245.

**Gonçalves-Ferreira, A. J., Herculano-Carvalho, M. and Pimentel, J.** (2003). Stereotactic biopsies of focal brainstem lesions. *Surgical Neurology* 60, 311–320.

**Lopes, A. P., Cardoso, L. and Rodrigues, M.** (2008). Serological survey of *Toxoplasma gondii* infection in domestic cats from northeastern Portugal. *Veterinary Parasitology* 155, 184–189.

**Lopes, A. P., Santos, H., Neto, F., Rodrigues, M., Kwok, O. C. H., Dubey, J. P. and Cardoso, L.** (2011a). Prevalence of antibodies to *Toxoplasma gondii* in dogs from northeastern Portugal. *Journal of Parasitology* 97, 418–420.

**Lopes, A. P., Sargo, R., Rodrigues, M., Cardoso, L.** (2011b). High seroprevalence of antibodies to *Toxoplasma gondii* in wild animals from Portugal. *Parasitology Research* 108, 1163–1169.

**Lopes, A. P., Dubey, J. P., Moutinho, O., Gargaté, M. J., Vilares, A., Rodrigues, M., Cardoso, L.** (2012). Seroepidemiology of *Toxoplasma gondii* infection in women from the North of Portugal in their childbearing years. *Epidemiology and Infection* 140, 872–877.

**Lopes, A. P., Dubey, J. P., Neto, F., Rodrigues, A., Martins, T., Rodrigues, M. and Cardoso, L.** (2013a). Seroprevalence of *Toxoplasma gondii* infection in cattle, sheep, goats and pigs from the North of Portugal for human consumption. *Veterinary Parasitology* 193, 266–269.

- Lopes, A. P., Sousa, S., Dubey, J. P., Ribeiro, A. J., Silvestre, R., Cotovio, M., Schallig, H. D., Cardoso, L. and Cordeiro-da-Silva, A.** (2013b). Prevalence of antibodies to *Leishmania infantum* and *Toxoplasma gondii* in horses from the north of Portugal. *Parasites and Vectors* **6**, 178.
- Miranda, A. M., Gomes, M. H., Guimarães, M., Nogueira, A., Abreu, C. and Figueiredo, P.** (1992). Toxoplasmose cerebral em doentes com SIDA. *Revista Portuguesa de Doenças Infecciosas* **15**, 163–172.
- Palmares, J., Coutinho, M. F. and Castro-Correia, J.** (1990). Uveitis in northern Portugal. *Current Eye Research* **9**(Suppl.), S31–S34.
- Robert-Gangneux, F. and Dardé, M. L.** (2012). Epidemiology of and diagnostic strategies for toxoplasmosis. *Clinical Microbiology Reviews* **25**, 264–296.
- Sousa, S., Ajzenberg, D., Vilanova, M., Costa, J. and Dardé, M. L.** (2008). Use of GRA6-derived synthetic polymorphic peptides in an immunoenzymatic assay to serotype *Toxoplasma gondii* in human serum samples collected from three continents. *Clinical and Vaccine Immunology* **15**, 80–386.
- Sousa, S., Thompson, G., Silva, E., Freire, L., Lopes, D., Correia da Costa, J. M., Castro, A., Carvalheira, J. and Canada, N.** (2009). Determination of the more adequate modified agglutination test cut-off for serodiagnosis of *Toxoplasma gondii* infection in sheep. *Zoonoses and Public Health* **56**, 252–256.
- Sousa, S., Canada, N., Correia da Costa, J. M. and Dardé, M. L.** (2010). Serotyping of naturally *Toxoplasma gondii* infected meat-producing animals. *Veterinary Parasitology* **169**, 24–28.
- Su, C., Khan, A., Zhou, P., Majumdar, D., Ajzenberg, D., Dardé, M. L., Zhu, X. Q., Ajioka, J. W., Rosenthal, B. M., Dubey, J. P. and Sibley, L. D.** (2012). Globally diverse *Toxoplasma gondii* isolates comprise six major clades originating from a small number of distinct ancestral lineages. *Proceedings of the National Academy of Science of the United States of America* **109**, 5844–5849.
- Waap, H., Vilares, A., Rebelo, E., Gomes, S. and Ângelo, H.** (2008). Epidemiological and genetic characterization of *Toxoplasma gondii* in urban pigeons from the area of Lisbon (Portugal). *Veterinary Parasitology* **157**, 306–309.
- Waap, H., Cardoso, R., Leitão, A., Nunes, T., Vilares, A., Gargaté, M. J., Meireles, J., Cortes, H. and Ângelo, H.** (2012). *In vitro* isolation and seroprevalence of *Toxoplasma gondii* in stray cats and pigeons in Lisbon, Portugal. *Veterinary Parasitology* **187**, 542–247.