

Asplenioideae Species as a Reservoir of Volatile Organic Compounds with Potential Therapeutic Properties

Didier Froissard, Sylvie Rapior, Jean-Marie Bessière, Bruno Buatois, Alain Fruchier, Vincent Sol, Françoise Fons

▶ To cite this version:

Didier Froissard, Sylvie Rapior, Jean-Marie Bessière, Bruno Buatois, Alain Fruchier, et al.. Asplenioideae Species as a Reservoir of Volatile Organic Compounds with Potential Therapeutic Properties. Natural Product Communications , 2015, 10 (6), pp.1079-1083. 10.1177/1934578X1501000671. hal-01257591

HAL Id: hal-01257591 https://unilim.hal.science/hal-01257591

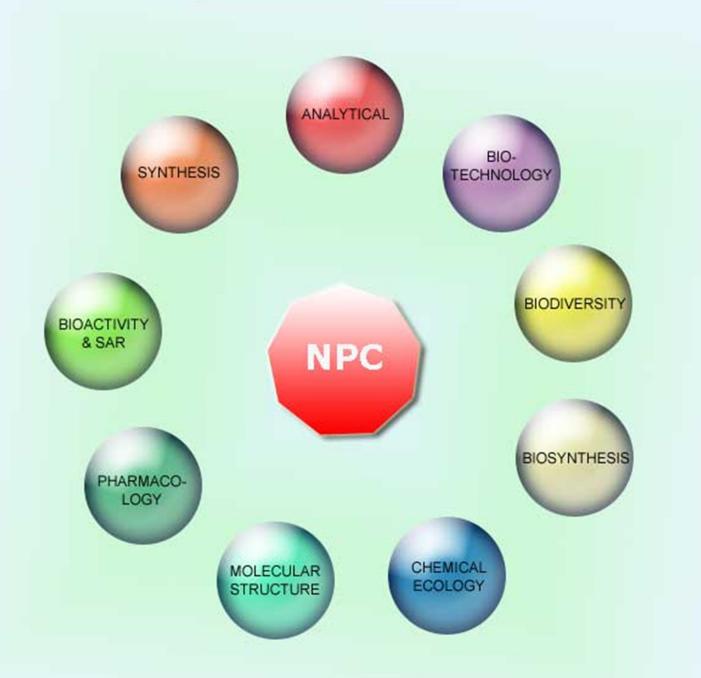
Submitted on 26 Jul 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

NATURAL PRODUCT COMMUNICATIONS

An International Journal for Communications and Reviews Covering all Aspects of Natural Products Research



This Issue is Dedicated to Dr. Pawan K. Agrawal On the Occasion of his 60th Birthday

Volume 10. Issue 6. Pages 823-1140. 2015 ISSN 1934-578X (printed); ISSN 1555-9475 (online) www.naturalproduct.us

NPC Natural P

Natural Product Communications

EDITOR-IN-CHIEF

DR. PAWAN K AGRAWAL

Natural Product Inc. 7963, Anderson Park Lane, Westerville, Ohio 43081, USA agrawal@naturalproduct.us

EDITORS

PROFESSOR ALEJANDRO F. BARRERO

Department of Organic Chemistry, University of Granada, Campus de Fuente Nueva, s/n, 18071, Granada, Spain afbarre@ugr.es

PROFESSOR ALESSANDRA BRACA

Dipartimento di Chimica Bioorganicae Biofarmacia, Universita di Pisa, via Bonanno 33, 56126 Pisa, Italy braca@farm.unipi.ti

PROFESSOR DE-AN GUO

State Key Laboratory of Natural and Biomimetic Drugs, School of Pharmaceutical Sciences, Peking University, Beijing 100083, China gda5958@163.com

PROFESSOR YOSHIHIRO MIMAKI

School of Pharmacy, Tokyo University of Pharmacy and Life Sciences, Horinouchi 1432-1, Hachioji, Tokyo 192-0392, Japan mimakiy@ps.toyaku.ac.jp

PROFESSOR STEPHEN G. PYNE

Department of Chemistry University of Wollongong Wollongong, New South Wales, 2522, Australia spyne@uow.edu.au

PROFESSOR MANFRED G. REINECKE

Department of Chemistry, Texas Christian University, Forts Worth, TX 76129, USA m.reinecke@tcu.edu

PROFESSOR WILLIAM N. SETZER

Department of Chemistry
The University of Alabama in Huntsville
Huntsville, AL 35809, USA
wsetzer@chemistry.uah.edu

PROFESSOR YASUHIRO TEZUKA

Faculty of Pharmaceutical Sciences Hokuriku University Ho-3 Kanagawa-machi, Kanazawa 920-1181, Japan y-tezuka@hokuriku-u.ac.jp

PROFESSOR DAVID E. THURSTON

Department of Pharmacy and Forensic Science, King's College London, Britannia House, 7 Trinity Street, London SEI 1DB, UK. david.thurston@kcl.ac.uk

HONORARY EDITOR

PROFESSOR GERALD BLUNDEN

The School of Pharmacy & Biomedical Sciences, University of Portsmouth, Portsmouth, PO1 2DT U.K. axuf64@dsl.pipex.com

ADVISORY BOARD

Prof. Viqar Uddin Ahmad Karachi, Pakistan Prof. Giovanni Appendino Novara, Italy Prof. Yoshinori Asakawa

Prof. Roberto G. S. Berlinck São Carlos, Brazil

Prof. Anna R. Bilia Florence, Italy Prof. Maurizio Bruno Palermo, Italy

Prof. César A. N. Catalán Tucumán, Argentina Prof. Josep Coll Barcelona, Spain Prof. Geoffrey Cordell Chicago, IL, USA Prof. Fatih Demirci Eskişehir, Turkey

Prof. Ana Cristina Figueiredo

Lisbon, Portugal

Prof. Cristina Gracia-Viguera Murcia, Spain

Dr. Christopher Gray Saint John, NB, Canada

Prof. Dominique Guillaume *Reims, France*Prof. Duvvuru Gunasekar

Tirupati, India

Prof. Hisahiro Hagiwara Niigata, Japan Prof. Tsukasa Iwashina Tsukuba, Japan Prof. Leopold Jirovetz Vienna, Austria Prof. Vladimir I Kalinin Vladivostok, Russia Prof. Phan Van Kiem Hanoi, Vietnam Prof. Niel A. Koorbanally Durban, South Africa Prof. Chiaki Kuroda Tokyo, Japan Prof. Hartmut Laatsch Gottingen, Germany

Prof. Marie Lacaille-Dubois

Dijon, France

Prof. Shopi Shopa Lee

Prof. Shoei-Sheng Lee Taipei, Taiwan Prof. Imre Mathe Szeged, Hungary

Prof. M. Soledade C. Pedras Saskatoon, Canada Prof. Luc Pieters Antwerp, Belgium Prof. Peter Proksch Düsseldorf, Germany

Prof. Phila Raharivelomanana Tahiti, French Polynesia Prof. Luca Rastrelli Fisciano, Italy Prof. Stefano Serra Milano, Italy

Prof. Monique Simmonds Richmond, UK

Dr. Bikram Singh Palampur, India Prof. John L. Sorensen Manitoba, Canada Prof. Johannes van Staden Scottsville, South Africa Prof. Valentin Stonik Vladivostok, Russia Prof. Winston F. Tinto

Prof. Sylvia Urban
Melbourne, Australia
Prof. Karen Valant-Vetschera

Vienna, Austria

INFORMATION FOR AUTHORS

Full details of how to submit a manuscript for publication in Natural Product Communications are given in Information for Authors on our Web site http://www.naturalproduct.us.

Authors may reproduce/republish portions of their published contribution without seeking permission from NPC, provided that any such republication is accompanied by an acknowledgment (original citation)-Reproduced by permission of Natural Product Communications. Any unauthorized reproduction, transmission or storage may result in either civil or criminal liability.

The publication of each of the articles contained herein is protected by copyright. Except as allowed under national "fair use" laws, copying is not permitted by any means or for any purpose, such as for distribution to any third party (whether by sale, loan, gift, or otherwise); as agent (express or implied) of any third party; for purposes of advertising or promotion; or to create collective or derivative works. Such permission requests, or other inquiries, should be addressed to the Natural Product Inc. (NPI). A photocopy license is available from the NPI for institutional subscribers that need to make multiple copies of single articles for internal study or research purposes.

To Subscribe: Natural Product Communications is a journal published monthly. 2015 subscription price: US\$2,595 (Print, ISSN# 1934-578X); US\$2,595 (Web edition, ISSN# 1555-9475); US\$2,995 (Print + single site online); US\$595 (Personal online). Orders should be addressed to Subscription Department, Natural Product Communications, Natural Product Inc., 7963 Anderson Park Lane, Westerville, Ohio 43081, USA. Subscriptions are renewed on an annual basis. Claims for nonreceipt of issues will be honored if made within three months of publication of the issue. All issues are dispatched by airmail throughout the world, excluding the USA and Canada.

Natural Product Communications

2015 Vol. 10 No. 6 1079 - 1083

Asplenioideae Species as a Reservoir of Volatile Organic Compounds with Potential Therapeutic Properties

Didier Froissard^a, Sylvie Rapior^b, Jean-Marie Bessière^c, Bruno Buatois^c, Alain Fruchier^d, Vincent Sol^a and Françoise Fons^b*

- ^a Laboratoire de Chimie des Substances Naturelles, LCSN, EA 1069, Faculté de Pharmacie de Limoges,
- 2 rue du Docteur Marcland, F-87025 Limoges Cedex, France
- ^b Laboratoire de Botanique, Phytochimie et Mycologie, Faculté de Pharmacie, CEFE UMR 5175, CNRS - Université de Montpellier - Université Paul-Valéry Montpellier – EPHE, 15 avenue Charles Flahault, F-34093 Montpellier Cedex 5, France
- ^c Centre d'Ecologie Fonctionnelle et Evolutive Plate-forme d'analyses chimiques en écologie, UMR 5175 CEFE, 1919 Route de Mende, F-34293 Montpellier Cedex 5, France
- ^d ENSCM, UMR 5253, 8 Rue de l'Ecole Normale, F-34296 Montpellier Cedex 5, France.

francoise.fons@univ-montp1.fr

Received: September 17th, 2014; Accepted: November 14th, 2014

Twelve French Asplenioideae ferns (genera Asplenium and subgenera Ceterach and Phyllitis) were investigated for the first time for volatile organic compounds (VOC) using GC-MS. Sixty-two VOC biosynthesized from the lipidic, shikimic, terpenic and carotenoid pathways were identified. Several VOC profiles can be highlighted from Asplenium jahandiezii and A. ×alternifolium with exclusively lipidic derivatives to A. onopteris with an equal ratio of lipidic/shikimic compounds. Very few terpenes as caryophyllene derivatives were identified, but only in A. obovatum subsp. bilotii. The main odorous lipidic derivatives were (E)-2-decenal (waxy and fatty odor), nonanal (aldehydic and waxy odor with a fresh green nuance), (E)-2-heptenal (green odor with a fatty note) and 1-octen-3-ol (mushroom-like odor), reported for all species. A few VOC are present in several species in high content, i.e., 9-oxononanoic acid used as a precursor for biopolymers (19% in A. jahandiezii), 4-hydroxyectophenone with a sweet and heavy floral odor (17.1% in A. onopteris), and 4-hydroxybenzoic acid used as a precursor in the synthesis of parabens (11.3% in A. foreziense). Most of the identified compounds have pharmacological activities, i.e., octanoic acid as antimicrobial, in particular against Salmonellas, with fatty and waxy odor (41.1% in A. petrarchae), tetradecanoic acid with trypanocidal activity (13.3% in A. obovatum subsp. bilotii), 4-hydroxybenzoic acid (8.7% in A. onopteris) with antimicrobial and anti-aging effects, 3,4-dihydroxybenzaldehyde as an inhibitor of growth of human cancer cells (6.7% in Ceterach officinarum), and phenylacetic acid with antifungal and antibacterial activities (5.8% in A. onopteris). Propionylfilicinic acid was identified in the twelve species. The broad spectrum of odorous and bioactive VOC identified from the Asplenium, Ceterach and Phyllitis species are indeed of great interest to the cosmetic and food industries.

Keywords: Asplenium, Ceterach, Phyllitis, Volatile Organic Compounds, 9-Oxononanoic acid, 3,4-Dihydroxybenzaldehyde, Antimicrobial, Anti-aging.

Asplenioideae Link is a great and homogeneous subfamily of Aspleniaceae Newman. Asplenium L. is the major genus with approximately seven hundred subcosmopolitan ferns distributed worldwide, and seventeen species in France. Several subgenera have been separated, such as Ceterach (Willd) Vida ex Bir, Fraser-Jenkins & Lovis and Phyllitis (Hill) Jermy & Viane [1a-1e].

The scientific name Asplenium was given by Pedanius Dioscorides (Roman physician, pharmacologist and botanist of Greek origin) to these plants that are well-known for their medicinal properties to cure the spleen; their common name "spleenwort" derives from the doctrine of signatures. Aspleniaceae includes many species reported for various traditional medicinal uses. Leaves and/or rhizomes of Asplenium adiantum-nigrum L., Ceterach officinarum Willd. (= A. ceterach L.), A. cuneatum Lam., A. falcatum Lam., A. marinum L., A. monanthes L., A. nidus L., A. ruta-muraria L., Phyllitis scolopendrium (L.) Newman (= A. scolopendrium L.) and A. trichomanes L. are used against worms, lung afflictions, cough inflammation, hypertension, jaundice, enlarged spleen, intestinal disorder, kidney stones, burns, elephantiasis and ulcers, and as an emetic, depurative, diaphoretic and sedative in traditional medicine [2a-2g]. Recently, antioxidant, antimicrobial and antibacterial properties of A. ceterach and A. nidus were demonstrated [2h-2i]. Regarding the chemical composition of Aspleniaceae, A. adiantum nigrum, A. fontanum, A. foresiense, A. incisum, A. normale, A.

obovatum, A. ruta-muraria, A. trichomanes and Ceterach officinarum were investigated for their phenolic derivatives [3a-3e].

Very few *Asplenium* are known to have an odor: *A. auritum* Sw. has pleasantly fragrant fronds and *A. lamprophyllum* Carse smells of wintergreen [4a]. Consequently, little is known about the volatile organic compounds (VOC) of these ferns. The terpenoid constituents of *A. scolopendrium* were studied [4b]. In addition, *A. trichomanes* subsp. *trichomanes* was investigated for its volatile profile [4c], which showed mainly polyketides, for example octanoic acid (= caprylic acid; fatty and waxy odor), nonanoic acid (waxy, dairy note), (*E*)-2-decenol (waxy note), (*E*)-2-heptenal (green odor with a fatty note) with globally an oily or waxy odor.

In this new work, fresh aerial parts of twelve French species of *Aspleniaceae* were investigated for their volatile profiles using GC-MS, as reported in the literature for the twenty-three monilophytes previously studied [4c-4f]. Sixty-two components biosynthesized from the shikimic, lipidic, terpenic and carotenoid pathways were identified from the concentrated diethyl ether extracts of the twelve *Aspleniaceae* (Table 1).

The VOC profile of *Ceterach officinarum* is widely dominated by lipidic derivatives (77.4%), in particular (*E*)-2-decenal (10.5%), a natural plant and mushroom VOC with waxy and fatty odor type

 Table 1: Percentage of volatile organic compounds^a in fresh aerial part of Asplenium, Ceterach and Phyllitis species.

Compounds	RIb	Ceterach officinarum s	Phyllitis scolopendriun	Asplenium n onopteris p	A. petrarchae	A. jahandiezi	A.obovatum i ssp. bilotii se	A. eptentrional	A. e foreziense	A. balearicun	A. ruta- n muraria	A. fontanum	A. ×alternifolium
Lipidic derivatives		77.4	79.3	46.9	71.3	98.0	59.4	79.9	66.0	89.9	74.5	83.6	99.3
Heptanal	905	0.6	• •		• 0	1.2	0.2			1.5	0.6	0.8	3.3
(E)-2-Heptenal 1-Octen-3-ol	957	8.6 5.7	3.9 8.8	5.8 5.8	2.8 2.0	10.5 3.7	3.9 4.5	3.1 2.7	6.7 1.8	13.0 7.4	5.6 1.7	8.3 3.9	8.8 6.1
2,3-Octanedione	982 986	0.5	1.9	3.8	2.0	2.9	4.3	0.7	1.8	1.5	1./	1.2	3.2
2-Pentylfuran	991	1.9	1.7	0.9	0.6	1.9	0.9	1.5	1.8	2.7	1.9	2.6	4.3
Octanal	1005	0.2	1.7	2.1	0.1	2.2	0.7	0.3	1.0	1.6	1.3	1.2	1.8
Hexanoic acid	1008	0.2	1.8	2.1	0.4	2.0		3.7		1.0	1.3	1.3	0.4
(Z)-3-Hexenoic acid	1017			1.1	0.1	0.3	0.5	0.7				0.2	1.1
3-Octen-2-one	1029	3.1		1.0	0.7	0.5	1.4	2.1		2.2		0.4	4.2
(E)-2-Hexenoic acid	1035			1.3	0.1	1.8	1.1	2.3			0.9	0.5	2.0
(E)-2-Octenal	1056	1.1		0.3	0.6	1.8		2.3		1.2	1.5	1.0	1.4
(E)-2-Octenol	1069	0.5		1.0	0.2	0.5	0.5	1.5		1.7		1.0	1.9
Octanol	1067		2.1	0.8	0.4	2.4	0.1			4.0	2.0	1.5	4.1
NI	1098	2.3	0.5	• •	• •			3.1					
Nonanal	1104	10.0	9.3	2.8	2.8	7.5	7.2	5.0	11.2	13.3	14.0	8.5	4.4
N-Acetylpyrrolidone (E)-2-Nonenal	1155 1162	2.9 1.0	1.2		0.5 0.4	0.7 0.8	0.5	3.1 0.7	0.9	0.9	1.3	1.4	1.6
Octanoic acid (Caprylic acid)	1185	1.1	2.2	2.8	41.1	5.4	1.8	3.2	7.2	1.2	3.3	3.9	3.2
(E,E)-2,4-Nonadienal	1215	1.0	2.2	0.5	0.2	1.7	0.4	0.7	1.2	0.4	1.1	0.8	0.9
N I	1228	0.5		0.5	0.2	0.4	0.4	0.7	1.2	0.7	1.1	0.0	0.7
NI	1256	0.3			0.3	0.1			3.3				
(E)-2-Decenal	1261	10.5	6.5	6.2	4.2	13.1	6.1	10.9	13.0	20.2	13.1	11.5	12.8
2-Decenol	1265	0.3	0.5		0.8	2.1	0.9	1.9		2.2	0.6	1.5	1.3
Nonanoic acid	1285	3.1	7.3	2.0	0.9	1.4	2.7			6.4	7.8	0.6	2.1
(E,Z)-2,4-Decadienal	1294		0.8	2.0	1.0	2.3	0.6		5.3	1.0		2.2	1.8
(E,E)-2,4-Decadienal	1319	2.7	2.0	3.1	1.5	2.2	1.1	3.1		2.0	1.3	3.2	4.1
NI	1361				0.5	2.5				1.6	2.6	0.9	0.7
Decanoic acid	1384	0.4	0.5		0.2	1.5		1.5	1.5	1.0	2.8	3.9	3.2
9-Oxononanoic acid	1502		6.2		2.1	19.0	2.2	3.7	3.8		4.4	4.1	7.0
N I	1510		7.9		0.5		0.7	4.6				1.5	0.2
Methyl laurate	1524	2.6	5.4	0.7	0.4	0.4	0.7	0.2	1.0	2.0	2.1	1.0	0.4
Propionylfilicinic acid	1552	3.6	5.4 2.0	0.7	0.4	0.4	1.2	0.3	1.8	2.8	2.1	1.8	0.4
Dodecanoic acid (Lauric acid)	1582	5.6	2.0			0.6	0.9	0.2				0.5	6.4
(E)-2-Tridecenoic acid N I	1687 1693	5.6 1.1	2.9	1.1		1.5	0.8	4.8 1.5				2.4	6.4 0.6
Methyl myristate	1726	1.1	2.9	1.1		1.3	1.8	1.3				1.0	0.0
Tetradecanoic acid	1778	4.8	2.8	1.8	1.4	3.0	13.3	3.7	6.5		3.3	7.3	4.2
Hexahydrofarnesylacetone	1840	4.8	1.1	2.5	1.2	0.3	2.7	5.6	0.5		5.5	2.2	1.2
Pentadecanoic acid	1885		***	1.0	1.2	0.5	1.7	1.5				0.5	1.8
N I	1917				2.3								
NI	1931				1.0								
Shikimic compounds		21.3	14.1	48.0	13.5	1.8	24.4	5.8	23.4	1.6	13.5	11.6	0
Benzaldehyde	961		2.3	1.0	0.4		0.6					1.2	
Benzyl alcohol	1043		1.4	1.1		0.8	1.6			1.6	1.3	1.4	
2-Phenylethanal	1046				0.7		0.2		1.5				
Benzoic acid	1195	1.0	3.6	2.1	4.6		1.7				1.1	0.5	
2,3-Dihydrobenzofuran	1235	0.2 2.3		2.0 0.7	1.5	1.0	1.1 2.7		0.9			1.2	
2-Amino-4-methoxyphenol Phenylacetic acid	1246 1265	0.4		5.8	1.3	1.0	0.7	1.9	0.9		5.6	1.2	
3,4-Dihydroxystyrene	1293	0.4		1.3	0.3		1.8	1.9	0.9		5.0		
Isovanillin	1397	0.2		1.5	0.3		0.8	0.4	1.8			1.5	
NI	1415			2.5	0.1		0.4	v	1.0		5.6	3.3	
Coumarin	1433							0.1					
4-Hydroxyacetophenone	1478			17.1									
Methyl 3-methoxy-4-													
hydroxybenzoate	1517		2.5	2.1	0.2		0.7		2.6				
4-Hydroxybenzoic acid	1560	5.8	3.5	8.7	4.2		7.8	0.3	11.3				
3-Methoxy-4-hydroxybenzoic				_			_					_	
acid	1568	4.8	0.8	0.7	1.5		4.4	2.1	4.4			2.5	
3,4-Dihydroxybenzaldehyde	1628	6.7		0.0				3.1					
3- or 4-Chlorocinnamic acid 3-Phenylpropionic acid	1638 1757			0.9 1.8									
Terpenic compounds	1/5/	0.0	0.0	0.0	5.8	0.0	2.3	0.0	0.0	0.0	0.0	0.0	0
Caryophyllene oxide	1586	0.0	0.0	0.0	3.0	0.0	0.8	0.0	0.0	0.0	0.0	0.0	U
β-Caryophyllene alcohol	1645						1.6						
N I	1683				1.6		1.0						
NI	1706				1.8								
NI	1760				2.4								
Carotenoid derivatives		0.8	5.9	4.3	9.0	0.0	13.1	13.3	9.7	7.6	11.4	4.0	0
α-Ionone	1435		2.1		0.2		1.3			2.0			
β-Ionone	1480			0.4	0.3		0.2						
Dihydroactinidiolide	1505		3.8	0.7	0.2		1.2	2.3	2.6			0.6	
4-Hydroxy-7,8-dihydro-β-													
ionone	1633						1.6			1.2			
4-Hydroxy-β-ionone	1639	0.3			4.6		1.9	1.8	1.5	0.6		1.8	
NI	1642				0.5		1.8						
3-Oxo-α-ionol	1647			1.2	0.7		1.3	0.2	1.8	2.0	2.1		
4-Hydroxy-5,6-epoxyionol	1675				2.3		1.7			1.8	7.8		
7,8-Epoxy-β-ionone	1688			1.4			1.3	0.5	3.8			0.7	
3-Hydroxy-5,6-epoxy-β-													
ionone	1696	0.2		0.5	0.2		1.1	3.1			1.5	0.9	
3-Oxo-7,8-dihydro-α-ionol	1710							3.1					
3-0x0-7,8-umyur0-a-101101								2.3					

^a Relative percentage of the VOC based on the GC-MS chromatographic area; ^bRI = Retention Indices on SLBTM-5MS column (Supelco); ^cNI = Not identified.

[5a-5c], nonanal (10%), with an aldehydic and waxy odor type and a fresh green nuance [5d] and recently reported to attract *Culex* mosquitoes [5e], (*E*)-2-heptenal (8.6%), with a green vegetable-like odor with fatty undertone, 1-octen-3-ol (5.7%), previously reported for mushrooms [5f, 5g], ferns [4c, 4d, 4f], horsetails [4e] and Angiosperms [5h], (*E*)-2-tridecenoic acid (5.6%) and hexahydrofarnesyl acetone (4.8%), which are used as flavor and fragrance agents. *C. officinarum* also contains several aromatic compounds (24.9%), i.e., 3,4-dihydroxybenzaldehyde (= protocatechuic aldehyde; 6.7%), a precursor of vanillin using biotechnology and recently reported to inhibit the growth of human cancer cells [5i], and 4-hydroxybenzoic acid (5.8%).

Phyllitis scolopendrium demonstrated a broad spectrum of VOC from the lipidic pathway (79.3%) including nonanal (9.3%), 1octen-3-ol (8.8%), a C₈-derivative responsible for the mushroomlike aroma [6a], nonanoic acid (7.3%), a C₉-derivative with waxy dairy note, (E)-2-decenal (6.5%), 9-oxononanoic acid (6.2%), (E)-2heptenal (3.9%), (E,Z)-dodecadienal (2.9%), tetradecanoic acid (myristic acid; 2.8%), octanoic acid (2.2%) and octanol (2.1%). It should be noted that propionylfilicinic acid (5.4%), a filicinic derivative of biological interest to the pharmaceutical, cosmetic and hygiene industries [4f], was identified in P. scolopendrium, as well as in the other ferns (Table 1). Benzoic acid, 4-hydroxybenzoic acid, methyl 3-methoxy-4-hydroxybenzoate (= methyl vanillate), with a warm spicy vanilla odor, and benzaldehyde, with a bitter almond odor [6b-6e], complete the aromatic profile (14.1%). Dihydroactinidiolide (3.8%), with a fruity odor [5d], and α -ionone (2.1%), with a floral smell, represent the major VOC of the carotenoid profile.

The VOC fraction of Asplenium onopteris is based on equal contents of shikimic and lipidic derivatives (48% and 46.9%, respectively). Twenty-one lipidic derivatives were identified, i.e., (E)-2-decenal (6.2%), (E)-2-heptenal (5.8%), 1-octen-3-ol (5.8%), (E,E)-2,4-decadienal (3.1%) with aldehyde, oily and fatty odor, and which is a potent plant nematicidal agent [7e]. The major aromatic compounds are 4-hydroxyacetophenone (17.1%), an aromatic ketone with a sweet, and heavy floral odor used for the synthesis of pharmaceuticals, agrochemicals, flavor and fragrances, and 4hydroxybenzoic acid (8.7%), used in cosmetic and ophthalmologic industries as a precursor in the synthesis of parabens, and which also shows antimicrobial activity and an anti-aging effect [7a, 7b], as well as a hypoglycemic property in rats [7c]. Phenylacetic acid (5.8%), used in the flavor industry for its honey-like odor, is a powerful antifungal and antibacterial agent, which is also produced by ants [7d]. Benzoic acid (2.1%), methyl 3-methoxy-4hydroxybenzoate (2.1%), 2,3-dihydrobenzofuran (2%), and 3phenylpropionic acid (= dihydrocinnamic acid) with a sweet, floral scent (1.8%), complete the shikimic derivatives. 7,8-Epoxy-βionone (1.4%) and 3-oxo- α -ionol (1.2%), with spicy odor, represent the main carotenoid derivatives (4.3%).

Asplenium petrarchae is a small and thermophile fern growing only on the sunny calcareous rocks of the Mediterranean coast. Its VOC profile is dominated by twenty-seven identified lipidic compounds (71.3%), with a very high level of octanoic acid (41.1%) with a fatty, soapy odor. This fatty acid is an antimicrobial ingredient used in particular against Salmonella species in cosmetics and foods; it presents the advantage of being less toxic than most other antimicrobial agents and does not affect beneficial organisms [8a-8c]. (E)-2-Decenal, (E)-2-heptenal, nonanal, 9-oxononanoic acid and 1-octen-3-ol were also identified as lipidic derivatives from A. petrarchae. The volatile fraction contains 13.5% of aromatic compounds, i.e., benzoic acid (4.6%), used as a food additive for its

preservative property, as well as 9% of carotenoid derivatives, i.e., 4-hydroxy-β-ionone (4.6%) and 4-hydroxy-5,6-epoxyionol (2.3%).

Asplenium jahandiezii is a small and protected fern only located in the canyon of Verdon (France) [1a]. Its VOC spectrum is almost exclusively dominated by twenty-nine identified lipidic compounds (98%). The major volatile is 9-oxononanoic acid (19%), an interesting VOC as a renewable resource of a precursor for biopolymers [9a]. It was recently discovered that 9-oxononanoic acid stimulates the activity of phospholipase A2, the key enzyme of the arachidonate cascade [9b]. The other lipidic derivatives with various odorous or pharmacological properties were (E)-2-decenal (13.1%), (E)-2-heptenal (10.5%), nonanal (7.2%), octanoic acid (5.4%), 1-octen-3-ol (3.7%) and tetradecanoic acid (3%). The minor odorous and bioactive lipidic derivatives were 2,3-octanedione (2.9%), with a dill cooked broccoli buttery odor [9c], and octanol (2.4%), with a sweetish odor and toxic to Colletotrichum gloeosporioides, an endophytic plant pathogen [9d], as well as (E,Z)-2,4-decadienal (2.3%), with a fatty, green and waxy odor, (E,E)-2,4-decadienal (2.2%), octanal (2.2%), with an aldehyde, fatty, orange peel, pungent and soapy flavor, (E)-2-decenol (2.1%), with a waxy, citrus and fresh note, and hexanoic acid (2%).

Asplenium obovatum subsp. bilotii has a broad spectrum of VOC with a high content of lipidic derivatives (59.4%) including 13.3% of tetradecanoic acid. This saturated fatty acid is one of the lipidic constituents of the cellular membrane of Eucaryotes. It is used as a lubricant and in the manufacture of flavors, pharmaceuticals, soaps and cosmetics, and has a trypanocidal activity, which was highlighted against Trypanosoma evansi two decades ago [10a]. This acid was identified in eleven of the studied ferns (Table 1). Nonanal (7.2%), (E)-2-decenal (6.1%), 1-octen-3-ol (4.5%), (E)-2heptenal (3.9%), nonanoic acid (2.7%), hexahydrofarnesylacetone (2.7%), 9-oxononanoic acid (2.2%), and octanoic acid (1.8%) were also identified from the organic extract. Aromatic compounds represent 25.5% of the VOC content of A. obovatum subsp. bilotii with 4-hydroxybenzoic acid (7.8%), 3-methoxy-4-hydroxybenzoic acid (4.4%), 2-amino-4-methoxyphenol (2.7%) as well as 3,4dihydroxystyrene (1.8%), an inhibitor of phenylalanine hydroxylase used for the production of experimental phenylketonuria [10b]. Finally, the carotenoids (13.1%) are dominated by 4-hydroxy-βionone (1.9%), 4-hydroxy-5,6-epoxyionol (1.7%) and 4-hydroxy-7,8-dihydro-β-ionone (1.6%), which is well-known as a key odorant in yellow wines [10c]. Two sesquiterpenes, i.e., caryophyllene alcohol and caryophyllene oxide, complete the VOC composition of A. obovatum subsp. bilotii. This oxygenated terpenoid, which is a flavoring agent used in cosmetics and food, also displays biological activities (anti-inflammatory, antifungal, skin enhancing and anticarcinogenic) [10d].

The six others Asplenium species also investigated for their VOC content for the first time were A. septentrionale, A. foreziense, A. balearicum, A. ruta-muraria, A. fontanum and Asplenium *alternifolium.

Most of their volatile constituents were mentioned above for the six first detailed VOC fern profiles. The main VOCs identified for A. septentrionale were two aldehydes, i.e., (E)-2-decenal (10.9%) and nonanal (5%). In addition A. septentrionale contained the highest concentration of hexahydrofarnesylacetone (5.6%), hexanoic acid (3.7%), a fatty acid found in animal oils with a fatty, waxy or cheesy flavor, N-acetylpyrrolidone (3.1%) and 3-hydroxy-5,6-epoxy- β -ionone (3.1%). A. foreziense also produced a high level of aldehydes (nonanal, (E)-2-decenal...) and the highest proportions of 4-hydroxybenzoic acid (11.3%), (E,Z)-2,4-decadienal (5.3%), with a

fatty, green and waxy odor, and 7,8-epoxy- β -ionone (3,8%) when compared with the other *Asplenioideae* (Table 1).

The highest percentages of (*E*)-2-decenal (20.2%), with a waxy fatty odor, and (*E*)-2-heptenal (13%), with a green fatty note, were found in *A. balearicum*, which also produced a high level of nonanal (13.3%), with an aldehydic and green scent. The volatile fraction of *A. ruta-muraria* contained 13.5% of shikimic compounds, i.e., phenylacetic acid (5.6%), as well as 11.4% of carotenoid derivatives, i.e., 4-hydroxy-5,6-epoxyionol (7.8%). Its major lipid derivatives (74.5%) were nonanal (14%), (*E*)-2-decenal (13.1%), nonanoic acid (7.8%) and (*E*)-2-heptenal (5.6%), already found in the other *Asplenium* species. The global VOC profile of this species (lipidic derivatives / shikimic derivatives ratio) can be compared with those of *P. scolopendrium* and *A. petrarchae*.

The major VOC of *A. fontanum* were three aldehydes, i.e., (*E*)-2-decenal, nonanal and (*E*)-2-heptenal, as well as tetradecanoic acid, already found in most of the *Asplenium* species. The *Asplenium* ×alternifolium VOC profile was close to that of *A. jahandiezii*, with uniquely lipidic derivatives (99.3%) and no shikimic compounds. The major lipidic compounds of *A.* ×alternifolium were (*E*)-2-decenal (12.8%), (*E*)-2-heptenal (8.8%), 9-oxononanoic acid (7%), (*E*)-2-tridecenoic acid (6.4%) and 1-octen-3-ol (6.1%), also produced by *C. officinarum* and *A. septentrionale*. Compared with the eleven other species investigated (Table 1), *A.* ×alternifolium contained the highest amount of 2-pentylfuran (4.3%), 3-octen-2-one (4.2%; earthy spicy herbal with mushroom nuances), octanol (4.1%), (*E,E*)-2,4-decadienal (4.1%), heptanal (3.3%), and 2,3-octanedione (3.2%).

Conclusion: The twelve French ferns from the family *Aspleniaceae* investigated for VOC mainly contain derivatives of lipidic origin. Several VOC profiles can be highlighted from A. *alternifolium, with exclusively lipidic derivatives, to A. onopteris, with an equal ratio of lipidic / shikimic compounds. Minor volatile components were identified from the shikimic pathway and very few terpenes, as caryophyllene derivatives, were found. Aldehydes, i.e., (E)-2-heptenal, nonanal and (E)-2-decenal, were often identified from the species, as well as acids (octanoic, nonanoic, 9-oxononanoic and tetradecanoic acids). 1-Octen-3-ol was found in all samples, as in most of the previously reported ferns [4c-4f]. It should be

mentioned that propionylfilicinic acid (from 0.3% to 5.4% of the volatile fraction) was identified in the twelve studied *Aspleniaceae*. This chemical trait must be noticed because many other ferns previously studied do not produce these volatile phloroglucinol derivatives; filicinic acids were mainly found in large amounts in *Dryopteris* [4f]. The broad spectrum of odorous and bioactive VOC identified from the twelve *Asplenium*, *Ceterach* and *Phyllitis* species from France are indeed of great interest for their various odorous and pharmacological properties that could be of interest to the cosmetics and food industries. Further investigations should be carried out through plant tissue cultures.

Experimental

Plant material: Fresh aerial parts of the ferns were collected as follows: C. officinarum: 13/07/2010, Limoges; P. scolopendrium: 13/07/2010, Limoges; A. onopteris: 05/04/2010, Le Pradet (Var); A. petrarchae: 15/04/2013, Toulon; A. jahandiezii: 26/07/2013, Aiguine (Var); A. obovatum subsp. bilotii: 14/04/2010, Le Lavandou (Var); A. septentrionale: 30/05/2011: Saint-Etienne-Vallée-Française (Lozère); A. foreziense: 20/09/2009, Meymac (Corrèze); A. balearicum: 16/07/2013, Porquerolle Island (Var); A. ruta-muraria: 13/07/2010, Limoges; A. fontanum: 20/07/2010 Plan d'Aups Sainte Baume (Var); Asplenium ×alternifolium: 30/05/2011: Saint-Etienne-Vallée-Française (Lozère). Authorization of harvest of the protected Aspleniaceae: 80-2013/06. Voucher specimens are deposited at the Laboratory of Botany (Faculty of Pharmacy, Limoges, France).

Plant part and GC-MS analyses: Fresh aerial parts of Asplenium, Ceterach and Phyllitis species were cubed and extracted with diethyl ether (Carlo Erba, 6 ppm BHT). After 1 week of maceration at room temperature, the concentrated organic extracts were used for gas chromatography mass spectrometry (GC-MS) analyses, as reported in the literature [4c-4f]. The main volatile components were identified by comparison with the National Institute of Standards and Technology Mass Spectral Library [10a-10b]. Internal standards (n-alkanes) were used as reference points in the calculation of relative retention indices. The analyses were performed at the Platform for Chemical Analyses in Ecology of the "SFR 119 Montpellier Environnement Biodiversité".

Acknowledgments - The authors greatly thank the Botanical Garden of Limoges for providing *Phyllitis scolopendrium*.

References

- [1] (a) Prelli R, Boudry M. (2001) Les fougères et plantes alliées de France et d'Europe occidentale, Belin, Paris, 1-432; (b) Pinter I, Bakker F, Barrett J, Cox C, Gibby M, Henderson S, Morgan-Richards M, Rumsey F, Russell S, Trewick S, Schneider H, Vogel J. (2002) Phylogenetic and biosystematic relationships in four highly disjunct polyploid complexes in the subgenera Ceterach and Phyllitis in Asplenium (Aspleniaceae). Organisms Diversity & Evolution, 2, 299-311; (c) Smith AR, Pryer KM, Schuettpelz E, Korall P, Schneider H, Wolf PG. (2006) A classification for extant ferns. Taxon, 55, 705-731; (d) Christenhusz M, Zhang XC, Schneider H. (2011) A linear sequence of extant families and genera of lycophytes and ferns. Phytotaxa, 19, 7-54; (e) Christenhusz MJM, Chase MW. (2014) Trends and concepts in fern classification. Annals of Botany, 113, 571-594.
- [2] (a) May LW. (1978) The economic uses and associated folklore of ferns and fern allies. The Botanical Review, 44, 491-528; (b) Guarrera PM, Salerno G, Caneva G. (2005) Folk phytotherapeutical plants from Maratea area (Basilicata, Italy). Journal of Ethnopharmacology, 99, 367-378; (c) Pieroni A, Dibra B, Grishaj G, Grishaj I, Macai SG. (2005) Traditional phytotherapy of the Albanians of Lepushe, Northern Albanian Alps. Fitoterapia, 76, 379–399; (d) Passalacqua NG, Guarrera PM, De Fine G. (2007) Contribution to the knowledge of the folk plant medicine in Calabria region (Southern Italy). Fitoterapia, 78, 52-68; (e) Mannan MM, Maridass M, Victor B. (2008) A review on the potential uses of ferns. Ethnobotanical Leaflets, 12, 281-285; (f) Irudayaraj V, Johnson M. (2011) Pharmacognostical studies on three Asplenium species. Journal of Phytology, 3, 1-9; (g) Benniamin A. (2011) Medicinal ferns of North Eastern India with special reference to Anapurna Pradesh. Indian Journal of Traditional Knowledge, 10, 516-522; (h) Berk S, Tepe B, Arslan S, Sarikurkcu C. (2011) Screening of the antioxidant, antimicrobial and DNA damage protection potentials of the aqueous extract of Asplenium ceterach DC. African Journal of Biotechnology, 10, 8902-8908; (i) Lai HY, Lim YY, Tan PT. (2009) Antioxidative, tyrosinase inhibiting and antibacterial activities of leaf extracts from medicinal ferns. Bioscience, Biotechnology, and Biochemistry, 73, 1362-1366.
- [3] (a) Đurđević L, Mitrović M, Pavlović P, Bojović S, Jarić S, Oberan L, Gajić G, Kostić O. (2007) Total phenolics and phenolic acids content in leaves, rhizomes and rhizosphere soil under *Ceterach officinarum* D.C., *Asplenium trichomanes* L. and *A. adiantum nigrum* L. in the Gorge of Sićevo (Serbia). *Ekologia (Bratislava)*, 26, 164-173; (b) Fan P, Zhao L, Hostetmann K, Lou H. (2012) Chemical constituents of *Asplenium ruta*-

- muraria L. Natural Product Research, 26, 1413-1418; (c) Imperato F. (1980) A new sulfated flavonol glycoside in the fern Asplenium fontanum Bernh. Chemistry & Industry, 13, 540-541; (d) Iwashina T, Matsumoto S, Ozawa K, Akuzawa K. (1990) Flavone glycosides from Asplenium normale. Phytochemistry, 29, 3543-3546; (e) Iwashina T, López-Sáez JA, Herrero A, Kitajima J, Matsumoto S. (2000) Flavone glycosides from Asplenium foreziense and its five related taxa and A. incisum. Biochemical Systematics and Ecology, 28, 665-671.
- [4] (a) Jones DL. (1987) Encyclopedia of ferns, Lothian, Port Melbourne, Australia; (b) Sohn YM, Chin YW, Yang MH, Kim J. (2008) Terpenoid constituents from the aerial parts of Asplenium scolopendrium. Natural Product Sciences, 14, 265-268; (c) Froissard D, Fons F, Bessière JM, Buatois B, Rapior S. (2011) Volatiles of French ferns and "fougère" scent in perfumery. Natural Product Communications, 6, 1723-1726; (d) Fons F, Froissard D, Bessière JM, Buatois B, Rapior S. (2010) Biodiversity of volatile organic compounds from five French ferns. Natural Product Communications, 5, 1655-1658; (e) Fons F, Froissard D, Bessière JM, Buatois B, Rapior S. (2013) Volatile organic compounds of six French horsetails. Natural Product Communications, 8, 509-512; (f) Froissard D, Fons F, Bessière JM, Fruchier A, Buatois B, Rapior S. (2014) Volatile organic compounds of six French Dryopteris species: Natural odorous and bioactive resources. Natural Product Communications, 6, 1723-1726.
- (a) Neff WE, Warner K, Byrdwell WC. (2000) Odor significance of undesirable degradation compounds in heated triolein and trilinolein. Journal of the American Oil Chemists' Society, 77, 1303-1313; (b) Xie J, Sun B, Zheng F, Xiao Y, Liu J. (2009) Supercritical CO2 fluid extraction of volatile flavor compounds from oxidized mutton fat. Shipin Kexue (Beijing, China), 30, 168-171; (c) Cho IH, Namgung HJ, Choi HK, Kim YS. (2007) Volatiles and key odorants in the pileus and stipe of pine-mushroom (Tricholoma matsutake Sing.). Food Chemistry, 106, 71-76; (d) Fons F, Rapior S, Eyssartier G, Bessière JM. (2003) Volatile compounds in the Cantharellus, Craterellus and Hydnum genera. Cryptogamie, Mycologie, 24, 367-376; (e) Syed Z, Leal WS. (2009) Acute olfactory response of Culex mosquitoes to a human- and bird-derived attractant. Proceedings of the National Academy of Sciences, 106, 18803-18808; (f) Rapior S, Breheret S, Talou T, Pélissier Y, Bessière JM. (2002) The anise-like odor of Clitocybe odora, Lentinellus cochleatus and Agaricus essettei. Mycologia, 94, 373-376; (g) Rapior S, Breheret S, Talou T, Pélissier Y, Milhau M, Bessière JM. (1998) Volatile components of fresh Agrocybe aegerita and Tricholoma sulfureum. Cryptogamie, Mycologie, 19, 15-23; (h) Fons F, Rapior S, Gargadennec A, Andary C, Bessière JM. (1998) Volatile components of Plantago lanceolata (Plantaginaceae). Acta Botanica Gallica, 145, 265-269; (i) Lee BH, Yoon SH, Kim YS, Kim SK, Moon BJ, Bae YS (2008) Apoptotic cell death through inhibition of protein kinase CKII activity by 3,4-dihydroxybenzaldehyde purified from Xanthium strumarium. Natural Product Research, 22, 1441-1450.
- (a) Talou T, Roule K, Gaset A. (1995) Arôme champignon: authentification par analyse chirale de l'octène-3-ol. Arômes Ingrédients Additifs, 3, 34-36; (b) Fons F, Rapior S, Fruchier A, Saviuc P, Bessière JM. (2006) Volatile composition of Clitocybe amoenolens, Tricholoma caligatum and Hebeloma radicosum. Cryptogamie, Mycologie, 27, 45-55; (c) Rapior S, Breheret S, Talou T, Pélissier Y, Bessière JM. (2002) The anise-like odor of Clitocybe odora, Lentinellus cochleatus and Agaricus essettei. Mycologia, 94, 373-376; (d) Rapior S, Fruchier A, Bessière JM. (1997) Volatile aroma constituents of agarics and boletes (A review, 134 references). In Recent Research Developments in Phytochemistry. Pandalai SG. (Ed). Research Signpost, Trivandrum, India, 1, 567-584; (e) Rapior S, Konska G, Guillot J, Andary C, Bessière JM. (2000) Volatile composition of Laetiporus sulphureus. Cryptogamie, Mycologie, 21, 67-72.
- (a) Kim KK, Jeon H, Cha DS. (2014) 4-Hydroxybenzoic acid-mediated lifespan extension in Caenorhabditis elegans. Journal of Functional Foods, 7, 630-640; (b) Cho JY, Moon JH, Seong KY, Park KH. (1998) Antimicrobial activity of 4-hydroxybenzoic acid and trans 4-hydroxycinnamic acid isolated and identified from rice hull. Bioscience, Biotechnology, and Biochemistry, 62, 2273-2276; (c) Peungvicha P, Temsiririrkkul R, Kurmar Prasain J, Tezuka Y, Kadota S, Thirawarapan SS, Watanabe H. (1998) 4-Hydroxybenzoic acid: a hypoglycemic constituent of aqueous extract of Pandanus odorus root. Journal of Ethnopharmacology, 62, 79-84; (d) Do Nascimento RR, Schoeters E, Morgan ED, Billen J, Stradling DJ. (1996) Chemistry of metapleural gland secretions of three attine ants, Atta sexdens rubropilosa, Atta cephalotes, and Acromyrmex octospinosus (Hymenoptera: Formicidae). Journal of Chemical Ecology, 22, 987-1000; (e) Caboni P, Ntalli NG, Aissani N, Cavoski I, Angioni A. (2012) Nematicidal activity of (E,E)-2,4-decadienal and (E)-2-decenal from Ailanthus altissima against Meloidogyne javanica. Journal of Agricultural and Food Chemistry, 60, 1146-1151.
- [8] (a) Harrison LM, Balan KV, Babu US. (2013) Dietary fatty acids and immune response to food-borne bacterial infections. *Nutrients*, 5, 1801-1822; (b) Moschonas G, Geornaras I, Stopforth JD, Wach D, Woerner DR, Belk KE, Smith, GC, Sofos JN. (2012) Activity of caprylic acid, carvacrol, epsilon-polylysine and their combinations against *Salmonella* in not-ready-to-eat surface-browned, frozen, breaded chicken products. *Journal of Food Science*, 77, 405-411; (c) Skrivanova E, Savka, OG, Marounek M. (2004) *In vitro* effect of C-2-C-18 fatty acids on salmonellas. *Folia Microbiologica*, 49, 199-202.
- [9] (a) Otte KB, Kirtz M, Nestl BM, Hauer B. (2013) Synthesis of 9-oxononanoic acid, a precursor for biopolymers. ChemSusChem, 6, 2149-2156; (b) Ren R, Hashimoto T, Mizuno M, Takigawa H, Yoshida M, Azuma T, Kanazawa K. (2013) A lipid peroxidation product 9-oxononanoic acid induces phospholipase A₂ activity and thromboxane A₂ production in human blood. Journal of Clinical Biochemistry and Nutrition, 52, 228-233; (c) Lipka C, Greenleaf S. (2011) Ingredient profile: 2,3-Octanedione. Perfumer & Flavorist, 36 (7), 22; (d) Nidiry ESJ. (2001) Structure–fungitoxicity relationships of some volatile flavour constituents of the edible mushrooms Agaricus bisporus and Pleurotus florida. Flavour and Fragrance Journal, 16, 245-248.
- (a) Ross CA, Taylor AM. (1994) Trypanocidal activity of a myristic acid analog in axenic cultures of *Trypanosoma evansi*. *Parasitology Research*, 80, 147-153; (b) Koizumi S, Matsushima Y, Nagatsu T, Iinuma H, Takeuchi T, Umezawa H.(1984) 3,4-Dihydroxystyrene, a novel microbial inhibitor for phenylalanine hydroxylase and other pteridine-dependent monooxygenases. *Biochimica et Biophysica Acta*, 789, 111-118; (c) Collin S, Nizet S, Claeys Bouuaert T, Despatures PM. (2012) Main odorants in Jura flor-sherry wines. Relative contributions of sotolon, abhexon, and theaspirane-derived compounds. *Journal of Agricultural and Food Chemistry*, 60, 380-387; (d) Yang D, Michel L, Chaumont JP, Millet-Clerc J. (1999) Use of caryophyllene oxide as an antifungal agent in an *in vitro* experimental model of onychomycosis. *Mycopathologia*, 148, 79-82.
- [11] (a) National Institute of Standard and Technology. (2005) PC version of the NIST / EPA / NIH Mass Spectra Database, Gaithersburg, Maryland, USA; (b) Adams RP. (2007) Identification of essential oil components by gas chromatography / mass spectroscopy. 4th Ed., Allured, Carol Stream, IL, USA.

A New Aromatic Compound from the Stem Bark of <i>Terminalia catappa</i> David Pertuit, Anne-Claire Mitaine-Offer, Tomofumi Miyamoto, Chiaki Tanaka, Stéphanie Delemasure, Patrick Dutartre and Marie-Aleth Lacaille-Dubois	1005
Effect of Non-psychotropic Plant-derived Cannabinoids on Bladder Contractility: Focus on Cannabigerol Ester Pagano, Vittorino Montanaro, Antonio di Girolamo, Antonio Pistone, Vincenzo Altieri, Jordan K. Zjawiony, Angelo A. Izzo and Raffaele Capasso	1009
In Cell Interactome of Oleocanthal, an Extra Virgin Olive Oil Bioactive Component Chiara Cassiano, Agostino Casapullo, Alessandra Tosco, Maria Chiara Monti and Raffaele Riccio	1013
Synthesis of & Viniferin Glycosides by Glucosyltransferase from <i>Phytolacca americana</i> and their Inhibitory Activity on Histamine Release from Rat Peritoneal Mast Cells Hiroki Hamada, Hatsuyuki Hamada and Kei Shimoda	1017
Stability of the Ellagitannin Fraction and Antioxidant Capacity of Varietal Pomegranate Juices Pedro Mena and Cristina García-Viguera	1017
Phthalide Anions in Organic Synthesis. A Direct Total Synthesis of Furomollugin George A. Kraus and Pengfei Dong	1025
Absolute Configuration Assignment of 3',4'-di-O-acylkhellactones Using Vibrational Circular Dichroism Exciton Chirality Abigail I. Buendía-Trujillo, J. Martín Torrres-Valencia, Pedro Joseph-Nathan and Eleuterio Burgueño-Tapia	1027
Antifouling Compounds from the Marine-Derived Fungus Aspergillus terreus SCSGAF0162 Xu-Hua Nong, Xiao-Yong Zhang, Xin-Ya Xu and Shu-Hua Qi	1033
Goji Berry: Quality Assessment and Crop Adaptation of Plants Cultivated in Tuscany (Italy) by Combination of Carotenoid and DNA Analyses	1033
Giada Capecchi, Emanuele Goti, Elena Nicolai, Maria Camilla Bergonzi, Roberto Monnanni and Anna Rita Bilia	1035
Activity of <i>Vitis vinifera</i> Tendrils Extract Against Phytopathogenic Fungi Daniele Fraternale, Donata Ricci, Giancarlo Verardo, Andrea Gorassini, Vilberto Stocchi and Piero Sestili	1037
Long-chain Glucosinolates from <i>Arabis turrita</i> : Enzymatic and Non-enzymatic Degradations Ivica Blažević, Sabine Montaut, Gina Rosalinda De Nicola and Patrick Rollin	1043
Aroma of Turmeric: Dependence on the Combination of Groups of Several Odor Constituents Toshio Hasegawa, Kenta Nakatani, Takashi Fujihara and Hideo Yamada	1047
Terpenoids Preserved in Fossils from Miocene-aged Japanese Conifer Wood Agnieszka Ludwiczuk and Yoshinori Asakawa	1051
Can Ozone Alter the Terpenoid Composition and Membrane Integrity of <i>in vitro Melissa officinalis</i> Shoots? Francesca D'Angiolillo, Mariagrazia Tonelli, Elisa Pellegrini, Cristina Nali, Giacomo Lorenzini, Luisa Pistelli and Laura Pistelli	1055
Composition and Chemical Variability of Ivoirian <i>Xylopia staudtii</i> Leaf Oil Thierry Acafou Yapi, Jean Brice Boti, Antoine Coffy Ahibo, Sylvain Sutour, Ange Bighelli, Joseph Casanova and Félix Tomi	1059
Chemoinformatics Approach to Antibacterial Studies of Essential Oils Dragoljub L. Miladinović, Budimir S. Ilić and Branislava D. Kocić	1063
Chemical Composition of Nardostachys grandiflora Rhizome Oil from Nepal – A Contribution to the Chemotaxonomy and Bioactivity of Nardostachys Prabodh Satyal, Bhuwan K. Chhetri, Noura S. Dosoky, Ambika Poudel and William N. Setzer	1067
Chemical Composition and Biological Activity of Essential Oils from Wild Growing Aromatic Plant Species of	
Skimmia laureola and Juniperus macropoda from Western Himalaya Iris Stappen, Nurhayat Tabanca, Abbas Ali, David E. Wedge, Jürgen Wanner, Vijay K. Kaul, Brij Lal, Vikas Jaitak, Velizar K. Gochev, Erich Schmidt and Leopold Jirovetz	1071
Comparative Chemical Composition and Antioxidant Properties of the Essential Oils of three Sideritis libanotica Subspecies Carmen Formisano, Filomena Oliviero, Daniela Rigano, Nelly Apostolides Arnold and Felice Senatore	1075
Asplenioideae Species as a Reservoir of Volatile Organic Compounds with Potential Therapeutic Properties Didier Froissard, Sylvie Rapior, Jean-Marie Bessière, Bruno Buatois, Alain Fruchier, Vincent Sol and Françoise Fons	1079
Composition and Comprehensive Antioxidant Activity of Ginger (Zingiber officinale) Essential Oil from Ecuador Martina Höferl, Ivanka Stoilova, Juergen Wanner, Erich Schmidt, Leopold Jirovetz, Dora Trifonova, Veselin Stanchev and Albert Krastanov	1085
Chemical Components of Four Essential Oils in Aromatherapy Recipe Sarin Tadtong, Narisa Kamkaen, Rith Watthanachaiyingcharoen and Nijsiri Ruangrungsi	1091
Accounts/Reivews	
Recent Advances in the Synthesis of Stemona Alkaloids Xiao-Yu Liu and Feng-Peng Wang	1093
Flavonoid Properties in Plant Families Synthesizing Betalain Pigments (Review) Tsukasa Iwashina	1103
Phytochemistry and Pharmacology of the Genus <i>Tovomita</i> Francesco Epifano, Maria Carmela Specchiulli, Vito Alessandro Taddeo, Serena Fiorito and Salvatore Genovese	1115
Fungal Phytotoxins with Potential Herbicidal Activity to Control Chenopodium album Alessio Cimmino, Marco Masi, Marco Evidente and Antonio Evidente	1119
Essential Oils as "A Cry for Help". A Review Christine Zitzelsberger and Gerhard Buchbauer	1127

Anti-Acetylcholinesterase Alkaloids from <i>Annona glabra</i> Leaf Shoei-Sheng Lee, Dong-Yi Wu, Sheng-Fa Tsai, and Chien-Kuang Chen	891
Increased Oxidative Stress in Cultured 3T3-L1 Cells was Attenuated by Berberine Treatment	
Shi-fen Dong, Naomi Yasui, Hiroko Negishi, Aya Kishimoto, Jian-ning Sun and Katsumi Ikeda Synthesis and Antimicrobial Activities of 3-Methyl-β-Carboline Derivatives	895
Jiwen Zhang, Longbo Li, Wenjia Dan, Jian Li, Qianliang Zhang, Hongjin Bai and Junru Wang	899
A Novel One-step Synthesis of Quinoline-2(1 <i>H</i>)-thiones and Selones by Treating 3-Aryl-3-(2-aminophenyl)-1-propyn-3-ols with a Base and Elemental Sulfur or Selenium	
Kazuaki Shimada, Hironori Izumi, Koki Otashiro, Kensuke Noro, Shigenobu Aoyagi, Yuji Takikawa and Toshinobu Korenaga	903
Normonanchocidins A, B and D, New Pentacyclic Guanidine Alkaloids from the Far-Eastern Marine Sponge <i>Monanchora pulchra</i> Ksenya M. Tabakmakher, Tatyana N. Makarieva, Vladimir A. Denisenko, Alla G. Guzii, Pavel S. Dmitrenok, Aleksandra S. Kuzmich and Valentin A. Stonik	913
Computational and Investigative Study of Flavonoids Active Against Trypanosoma cruzi and Leishmania spp	
Frederico F. Ribeiro, Francisco J.B.M. Junior, Marcelo S. da Silva, Marcus Tullius Scotti and Luciana Scotti Two New Secondary Metabolites from <i>Tephrosia purpurea</i>	917
Yin-Ning Chen, Yan Peng, Cheng-Hai Gao, Tao Yan, Zhi-Fang Xu, Samuel X. Qiu, Wen-Hao Cao, Ligao Deng and Ri-Ming Huang	921
Regioselective Glycosylation of 3-, 5-, 6-, and 7-Hydroxyflavones by Cultured Plant Cells Kei Shimoda, Naoji Kubota, Daisuke Uesugi, Yuuya Fujitaka, Shouta Okada, Masato Tanigawa and Hiroki Hamada	923
Unusual Flavonoid Glycosides from the Hawaiian Tree <i>Metrosideros polymorpha</i> Benjamin R. Clark, Swapan Pramanick, Norman Arancon and Robert P. Borris	925
Anti-inflammatory Flavonoids Isolated from <i>Passiflora foetida</i> Thi Yen Nguyen, Dao Cuong To, Manh Hung Tran, Joo Sang Lee, Jeong Hyung Lee, Jeong Ah Kim, Mi Hee Woo and Byung Sun Min	929
Clovamide and Flavonoids from Leaves of <i>Trifolium pratense</i> and <i>T. pratense</i> subsp. <i>nivale</i> Grown in Italy	729
Aldo Tava, Łukasz Pecio, Anna Stochmal and Luciano Pecetti	933
Water Extract of <i>Mentha</i> × <i>villosa</i> : Phenolic Fingerprint and Effect on Ischemia-Reperfusion Injury Silvia Fialova, Lucia Veizerova, Viera Nosalova, Katarina Drabikova, Daniela Tekelova, Daniel Grancai and Ruzena Sotnikova	937
Distribution and Taxonomic Significance of Secondary Metabolites Occurring in the Methanol Extracts of the Stonecrops (Sedum L., Crassulaceae) from the Central Balkan Peninsula Gordana S. Stojanović, Snežana Č. Jovanović and Bojan K. Zlatković	941
In vitro Xanthine Oxidase Inhibitory Studies of Lippia nodiflora and Isolated Flavonoids and Phenylethanoid Glycosides	
as Potential Uric Acid-lowering Agents Lee-Chuen Cheng, Vikneswaran Murugaiyah and Kit-Lam Chan	945
Enzymatic Synthesis of Quercetin Monoglucopyranoside and Maltooligosaccharides Ryo Yasukawa, Natsumi Moriwaki, Daisuke Uesugi, Fuya Kaneko, Hiroki Hamada and Shin-ichi Ozaki	949
Polyurethane Microstructures-a Good or Bad <i>in vitro</i> Partner for the Isoflavone Genistein? Corina Danciu, Florin Borcan, Codruta Soica, Istvan Zupko, Erzsébet Csányi, Rita Ambrus, Delia Muntean, Camelia Sass, Diana Antal, Claudia Toma and Cristina Dehelean	951
Chemical Constituents of the Underground Parts of <i>Iris florentina</i> and their Cytotoxic Activity Akihito Yokosuka, Yoshikazu Koyama and Yoshihiro Mimaki	955
Synthesis of Arecatannin A1 from Dimeric Epicatechin Electrophile Manato Suda, Kohki Takanashi, Miyuki Katoh, Kiriko Matsumoto, Koichiro Kawaguchi, Sei-ichi Kawahara, Hiroshi Fujii and Hidefumi Makabe	959
Anthocyanin Profile and Antioxidant Activity of Various Berries Cultivated in Korea	
Hong-Sook Bae, Hyun Ju Kim, Jin Hee Kang, Rika Kudo, Takahiro Hosoya, Shigenori Kumazawa, Mira Jun, Oh-Yoen Kim and Mok-Ryeon Ahn	963
Metabolite Fingerprinting of <i>Eugenia jambolana</i> Fruit Pulp Extracts using NMR, HPLC-PDA-MS, GC-MS, MALDI-TOF-MS and ESI-MS/MS Spectrometry	
Ram Jee Sharma, Ramesh C. Gupta, Arvind Kumar Bansal and Inder Pal Singh	969
Flavonoids and Phenolic Acids in Methanolic Extracts, Infusions and Tinctures from Commercial Samples of Lemon Balm Agnieszka Arceusz, Marek Wesolowski and Beata Ulewicz-Magulska	977
RP-HPLC-DAD-MS ⁿ Analysis and Butyrylcholinesterase Inhibitory Activity of Barbacenia blanchetii Extracts	
Jósquia S Barbosa, Verônica M Almeida, Rosilene M Marçal and Alexsandro Branco	983
Flavonoids and Other Phenolic Compounds in Needles of <i>Pinus peuce</i> and Other Pine Species from the Macedonian Flora Marija Karapandzova, Gjose Stefkov, Ivana Cvetkovikj, Jasmina Petreska Stanoeva, Marina Stefova and Svetlana Kulevanova	987
Anti-inflammatory, Antioxidant and Antimicrobial Activity Characterization and Toxicity Studies of Flowers of "Jarilla", a Medicinal Shrub from Argentina Alejandra Moreno, Gabriela Nuño, Soledad Cuello, Jorge E. Sayago, María Rosa Alberto, Catiana Zampini and María Inés Isla	991
Synthesis of Resveratrol Glycosides by Plant Glucosyltransferase and Cyclodextrin Glucanotransferase and Their	
Neuroprotective Activity Kei Shimoda, Naoji Kubota, Hatsuyuki Hamada and Hiroki Hamada	995
Anti-austeritic Constituents of the Congolese Medicinal Plant Aframomum melegueta Dya Fita Dibwe, Suresh Awale, Hiroyuki Morita and Yasuhiro Tezuka	997
The Lignan-containing Extract of Schisandra chinensis Berries Inhibits the Growth of Chlamydia pneumoniae	771
Elina Hakala, Leena L. Hanski, Teijo Yrjönen, Heikki J. Vuorela and Pia M. Vuorela	1001

Natural Product Communications 2015

Volume 10, Number 6

Contents

<u>Editorial</u>	i
<u>Preface</u>	iii
Original Paper	
Chemical and Genetic Diversity of <i>Ligularia hodgsonii</i> in China Chiaki Kuroda, Kou Inagaki, Xun Chao, Kyosuke Inoue, Yasuko Okamoto, Motoo Tori, Xun Gong, and Ryo Hanai	823
Constituents of <i>Ligularia brassicoides</i> Collected in China: A New Diels-Alder Adduct of Eremophilan-10β-ol and Methacrylic Acid Mizuho Taniguchi, Katsuyuki Nakashima, Yasuko Okamoto, Xun Gong, Chiaki Kuroda and Motoo Tori	827
Four New Sesquiterpenoids from Ligularia subspicata Collected in China; Isolation of a Bakkane-type Lactone, an Eremophilane-type Lactone, and Two Ortho Esters Yoshinori Saito, Takanori Otsubo, Yuko Iwamoto, Katsuyuki Nakashima, Yasuko Okamoto, Xun Gong, Chiaki Kuroda and Motoo Tori	831
Natural Caryophyllane Sesquiterpenoids from <i>Rumphella antipathies</i> Hsu-Ming Chung, Wei-Hsien Wang, Tsong-Long Hwang, Yang-Chang Wu and Ping-Jyun Sung	835
Bioactive Compounds in Wild, <i>In vitro</i> Obtained, <i>Ex vitro</i> Adapted, and Acclimated Plants of <i>Centaurea davidovii</i> (Asteraceae) Antoaneta Trendafilova, Milka Jadranin, Rossen Gorgorov and Marina Stanilova	839
New Laurene-type Sesquiterpene from Bornean <i>Laurencia nangii</i> Takashi Kamada and Charles Santhanaraju Vairappan	843
New Furanone and Sesquiterpene from the Pericarp of Calocedrus formosana Tzong-Huei Lee, Ming-Shian Lee, Horng-Huey Ko, Jih-Jung Chen, Hsun-Shuo Chang, Mei-Hwei Tseng, Sheng-Yang Wang, ODIVERS Chien-Chih Chen and Yueh-Hsiung Kuo	845
The Importance of the 5-Alkyl Substituent for the Violet Smell of Ionones: Synthesis of Racemic 5-Demethyl-α-ionone Serena Chierici, Serena Bugoni, Alessio Porta, Giuseppe Zanoni and Giovanni Vidari	847
Antiproliferative Activity of seco-Oxacassanes from Acacia schaffneri J. Martín Torres-Valencia, Virginia Motilva, J. Jesús Manríquez-Torres, Sofia García-Mauriño, Miguel López-Lázaro, Hanaa Zbakh, José M. Calderón-Montaño, Mario A. Gómez-Hurtado, Juan A. Gayosso-De-Lucio, Carlos M. Cerda-García-Rojas and Pedro Joseph-Nathan	853
neo-Clerodane Diterpenoids from Ajuga macrosperma var. breviflora Amaya Castro, Josep Coll, Anil K. Pant and Om Pakrash	857
Three New C ₂₀ -Diterpenoid Alkaloids from <i>Aconitum tanguticum</i> var. <i>trichocarpum</i> Zhong-Tang Zhang, Xiao-Yu Liu, Dong-Lin Chen, and Feng-Peng Wang	_861
Manoalide-related Sesterterpene from the Marine Sponge Luffariella variabilis Toshiyuki Hamada, Daisuke Harada, Mitsunobu Hirata, Keisuke Yamashita, Kishneth Palaniveloo, Hiroaki Okamura, Tetsuo Iwagawa, Naomichi Arima, Toshiyuki Iriguchi, Nicole J. de Voogd and Charles S. Vairappan	863
Oxygenated Terpenes from Indo-Pacific Nudibranchs: Scalarane Sesterterpenes from Glossodoris hikuerensis and 12-Acetoxy Dendrillolide A from Goniobranchus albonares I. Wayan Mudianta, Andrew M. White and Mary J. Garson	865
Germinating Seeds of Citrus aurantium a Good Source of Bioactive Limonoids Marta R. Ariza, M. Mar Herrador del Pino and Alejandro F. Barrero	869
Chemical Constituents of <i>Lecythis pisonis</i> (Lecythidaceae) – A New Saponin and Complete ¹ H and ¹³ C Chemical Shift Assignments Rennê C. Duarte, Carlos R. R. Matos, Raimundo Braz-Filho and Leda Mathias	871
Oleanane-type Triterpene Saponins from Glochidion glomerulatum Vu Kim Thu, Nguyen Van Thang, Nguyen Xuan Nhiem, Hoang Le Tuan Anh, Pham Hai Yen, Chau Van Minh, Phan Van Kiem, Nan Young Kim, Seon Ju Park and Seung Hyun Kim	875
Cucumarioside E from the Far Eastern Sea Cucumber Cucumaria japonica (Cucumariidae, Dendrochirotida), New Minor Monosulfated Holostane Triterpene Pentaoside with Glucose as the Second Monosaccharide Residue Alexandra S. Silchenko, Anatoly I.Kalinovsky, Pavel S. Dmitrenok, Vladimir I. Kalinin, Andrey N. Mazeika, Natalia S. Vorobieva, Nina M. Sanina and Edward Y. Kostetsky	877
Structure Revision of Two Polyoxygenated Sterols from the Marine Sponge Neofibularia nolitangere Yasunori Yaoita, Masao Kikuchi and Koichi Machida	881
Ergosterol of Cordyceps militaris Attenuates LPS Induced Inflammation in BV2 Microglia Cells Neeranjini Nallathamby, Lee Guan-Serm, Sharmili Vidyadaran, Sri Nurestri Abd Malek, Jegadeesh Raman and Vikineswary Sabaratnam	885
Two Novel Spirostene Glycosides from Selaginella chrysocaulos and their Chemotaxonomic Significance Olaf Kunert, Rumalla Chidananda Swamy, Bobbala Ravi Kumar, Achanta Venkata Narasimha Appa Rao, Owi Ivar Nandi and Wolfgang Schuehly	887