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Shining a Light on Prostate Cancer: Photodynamic Therapy and Combination Approaches

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Abstract: Prostate cancer is a major health concern worldwide, and current treatments, such as surgery, radiation therapy, and chemotherapy, are associated with significant side effects and limitations. Photodynamic therapy (PDT) is a promising alternative that has the potential to provide a minimally invasive and highly targeted approach to treating prostate cancer. PDT involves the use of photosensitizers (PSs) that are activated by light to produce reactive oxygen species (ROS), which can induce tumor cell death. There are two main types of PSs: synthetic and natural. Synthetic PSs are classified into four generations based on their structural and photophysical properties, while natural PSs are derived from plant and bacterial sources. Combining PDT with other therapies, such as photothermal therapy (PTT), photoimmunotherapy (PIT), and chemotherapy (CT), is also being explored as a way to improve its efficacy. This review provides an overview of conventional treatments for prostate cancer, the underlying principles of PDT, and the different types of PSs used in PDT as well as ongoing clinical studies. It also discusses the various forms of combination therapy being explored in the context of PDT for prostate cancer, as well as the challenges and opportunities associated with this approach. Overall, PDT has the potential to provide a more effective and less invasive treatment option for prostate cancer, and ongoing research is aimed at improving its selectivity and efficacy in clinical settings.

Keywords: photodynamic therapy; prostate cancer; reactive oxygen species; cell death; immune response; tumor vascular damage; clinical trials; combination approaches

1. Introduction

With over 1.4 million new cases and more than 375,000 deaths worldwide in 2020, prostate cancer is a major public health concern [1]. Prostate cancer is the second most commonly occurring cancer in men and the fourth most common cancer overall. In addition, the rising incidences of prostate cancer in the last decade are concerning, even if it is likely attributed to earlier detection through increased screening [1]. Conventional treatment options such as surgery, radiation therapy, hormone therapy, chemotherapy, and immunotherapy have limitations, including high invasiveness, poor specificity, and adverse effects. In this context, PDT has emerged as an innovative and promising treatment option for prostate cancer due to its high specificity, low invasiveness, and reproducibility [2,3].

PDT involves the use of photosensitizing agents and light to activate them, leading to localized cancer cell death [2]. However, PDT alone may not be sufficient to eradicate prostate cancer completely. Thus, the combination with other therapeutic options may pro-



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Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). vide opportunities to draw on the strong points of each option and offset their weaknesses, leading to additive or synergistic therapeutic effects [4].

This review article aims to explore the potential of PDT as a treatment for prostate cancer, including its mechanism of action, limitations, photosensitizers, and combination with other therapies. It also provides insights into the future directions of PDT in the context of prostate cancer treatment.

To ensure the accuracy and comprehensiveness of our review, our team embarked on a rigorous data collection and search process from a wide range of reputable databases, including Google Scholar, Pubmed, Springer, Elsevier ScienceDirect, and Web of Science. We focused on scrutinizing studies published from 2000 to 2023 and we included one older study for contextualization. In order to maintain consistency and ensure a thorough analysis, only articles containing English texts were selected, and we conducted keyword and heading searches for the following terms: prostate cancer, photodynamic therapy, natural photosensitizers, synthetic photosensitizers, prostate cancer treatment complications, and combination cancer therapies.

Then, a thorough selection process was conducted, starting with 2101 records. Before proceeding to screen full-text documents, duplicate papers and irrelevant works were eliminated. The inclusion criteria were strict and consisted of original articles, review papers, book chapters or websites that met specific factors, thus ensuring the accuracy and quality of the information presented in this paper. Overall, 96 references were selected for our study, which included articles or websites specifically dedicated to providing context, epidemiological data, and reports of clinical studies.

2. Conventional Treatments for Prostate Cancer and Complications

2.1. Surgery

Surgery is a common treatment option for prostate cancer and involves the partial or radical removal of the prostate gland. This procedure, known as a prostatectomy, can be performed using various techniques [5]. Nevertheless, many potential complications may arise in this procedure, such as incontinence and erectile dysfunction resulting from surgical damage to the urinary sphincter and erectile nerves [6]. These adverse effects can greatly impact a patient's quality of life and are important considerations in prostate cancer treatment decision-making [6].

2.2. Radiation Therapy

Radiation therapy is a cancer treatment that uses high-energy X-rays or other types of radiation to eliminate cancer cells. Two types of radiation therapy are used: external and internal [5]. External radiation therapy involves using a machine to direct radiation toward the cancer site, while internal radiation therapy places a radioactive substance directly into or near the cancer site [5,7]. Radiation therapy for prostate cancer increases the risk of developing bladder and/or gastrointestinal cancer. Furthermore, a higher rate of long-term complications, such as impotence and urinary problems, can also occur [5,8].

2.3. Hormone Therapy

Hormone therapy, also known as androgen deprivation therapy (ADT), is a treatment that aims to remove or block male hormones, which can cause cancer cells to grow. This can be achieved through drugs such as abiraterone acetate and luteinizing hormone-releasing hormone agonists, or surgery to remove the testicles (orchiectomy), or the use of antiandrogens that block the action of male hormones [5]. Hormone therapy can help shrink the tumor and slow the growth of cancer, but it may also cause side effects including hot flashes, loss of libido, and weakened bones [5,7].

2.4. Chemotherapy

Chemotherapy is a treatment that uses powerful drugs to destroy cancer cells. In prostate cancer, chemotherapy is usually reserved for advanced cases that no longer re-

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spond to hormone therapy [5]. Chemotherapy can cause side effects such as hair loss, nausea, vomiting, fatigue, and an increased risk of infection due to a weakened immune system. However, these side effects can be managed with medications and other supportive therapies [7,9].

2.5. Cellular Immunotherapy

Immunotherapy is a cancer treatment that harnesses the patient's own immune system to attack cancer cells. It involves the use of substances or cells, either naturally produced by the body or created in a lab, to enhance or restore the body's natural defenses against cancer [5]. In the case of prostate cancer, Sipuleucel-T, an autologous cellular immunological agent, is intended for patients with metastatic hormone-refractory prostate cancer. It consists of exposing the patient's peripheral blood mononuclear cells to a recombinant fusion protein composed of prostatic acid phosphatase, an antigen that is highly expressed in most prostate cancer cells, and a granulocyte-macrophage colony-stimulating factor. The mechanism of action is not fully elucidated, but it is likely to work through antigenpresenting cells (APCs) to stimulate a T-cell immune response [10]. The most reported side effects, including chills, fever, headache, myalgia, sweating, and influenza-like symptoms, were attributed to cytokine release usually within the first 24 h of infusion. However, no increased risk of autoimmune complications has been reported to date [5].

2.6. Other Therapeutic Approaches

Prostate cancer treatment has come a long way, with several innovative therapies reported in clinical studies [5]. Cryosurgery, or cryotherapy, involves the use of an instrument to freeze and destroy cancerous prostate cells. High-intensity-focused ultrasound therapy, on the other hand, uses ultrasound waves to break down cancer cells. Additionally, proton beam radiation therapy employs tiny, positively charged particles to target and kill tumor cells [5].

Besides these therapeutic approaches, another promising treatment for prostate cancer is PDT. The principle of this alternative treatment is the use of a photoactivated drug to selectively destroy cancer cells while minimizing damage to healthy tissues and side effects.

3. PDT

3.1. Mechanism of Action

PDT uses a photosensitizer (PS), molecular oxygen, and light to generate reactive oxygen species (ROS) that can trigger tumor cell death [11]. PDT can cause tumor destruction by three mechanisms: direct cell death, tumor vascular damage, and an immune response (Figure 1) [11,12].

Direct cell death (Figure 1A) occurs through both programmed cell death (apoptosis) and non-programmed cell death (necrosis) pathways [13]. Necrosis is characterized by rapid cell death leading to the release of cellular components and molecules that promote inflammation, while apoptosis is a genetically encoded, energy-dependent process that can initiate an immune response [14,15]. PDT can also induce unconventional modes of cell death in cancer cells, including paraptosis, parthanatos, mitotic catastrophe, pyroptosis, necroptosis, and ferroptosis [13]. In addition, a growing body of evidence suggests that PDT can also trigger immunogenic cell death (ICD) [16], which has emerged as a promising strategy for eliminating tumor cells by promoting T-cell adaptive immune responses and inducing durable immunological memory [13,16,17].

PDT can also damage the tumor microvasculature (Figure 1B), leading to the interruption of the tumor's feeding and, consequently, to the death of the tumor cells. This vascular mechanism is achieved by concentrating the PS in the vascular system and using a short drug-light interval [18]. PDT vascular effect can be selectively applied to the tumor and surrounding healthy tissue, with important advantages over PDT protocols that require PS accumulation in the tumor cells [17,18]. Finally, PDT can induce an immune response (Figure 1C) that can contribute to longterm tumor control [18]. PDT has been shown to influence the adaptive immune response through either stimulation or suppression, depending on the treatment protocol [6]. The oxidative damage inflicted by PDT on the tumor stroma triggers an acute inflammatory response initiated by the release of pro-inflammatory mediators. These mediators attract the host's innate immune cells, which can activate a systemic antitumor immune response. PDT-induced necrosis of tumors and their vasculature can activate CD8 cytotoxic T lymphocytes that can specifically destroy tumor cells and circulate throughout the body for long periods [18].



Figure 1. PDT destroys tumor cells by three mechanisms of action. **(A)** Induction of cell death through various pathways depends on the location of the photosensitizer (PS) and the extent of damage to the organelles. Mitochondrial damage caused by PDT results in the loss of membrane permeability and the release of pro-apoptotic mediators, while damage to the endoplasmic reticulum (ER) leads to the release of cellular calcium deposits. Lysosomal damage, on the other hand, causes the release of proteolytic enzymes upon illumination, and can also trigger autophagy. In cases where apoptosis is dysfunctional, necrosis and autophagy may be the primary modes of cell death after PDT. It is important to note that multiple PSs may localize in multiple organelles, and therefore, simultaneous activation of death pathways may occur (adapted from Mroz, P et al. [19]). **(B)** PDT can induce vascular damage, leading to the interruption of the tumor's feeding causing its death through various mechanisms. **(C)** PDT can also activate innate and adaptative immunity, leading to a systemic antitumor immune response and the destruction of tumor cells.

PSs are molecules that can absorb light and transfer the absorbed energy to the surrounding molecules (e.g., oxygen), leading to the generation of reactive oxygen species (ROS) that can cause cell death. These PSs are generally divided into two categories: tetrapyrrolic macrocycles (i.e., porphyrins, chlorins, bacteriochlorins, phthalocyanines, and their derivates) and no tetrapyrrolic macrocycles (i.e., methylene blue, curcumin, phenalenon, coumarin, anthraquinone, boron-dipyrromethene florophore (bodipy), and their derivatives) [20–23]. Tetrapyrrolic macrocycles are the most widely used PSs in anticancer PDT and, among them, porphyrins are the most studied. Thus, porphyrinderived PSs are further classified into three generations [24].

The first-generation PS, such as the hematoporphyrin derivative (HpD) and Photofrin[®], were the first photosensitizers to be used clinically in PDT. However, these molecules have some drawbacks, such as a mixture of PSs, prolonged skin photosensitization, no selectivity, and suboptimal tissue penetration [25].

Second-generation PSs were developed to address these issues. These compounds are chemically pure, absorb light at a longer wavelength, and cause significantly less skin photosensitization post-treatment. They must also be at least as efficient in eradicating tumors as Photofrin[®], which is currently the gold standard for PDT [26].

Third-generation PSs, which are bound to targeting agents (antibodies, sugar, folic acid, polyamine, peptide, etc.), are able to be selectively accumulated within tumor tissues and represent an active research area in the field [27–35].

Of note, fourth-generation PSs are still in the early stages of development and are characterized by their ability to be associated with nanoparticles and/or activated by specific stimuli, such as light at a specific wavelength, changes in pH or temperature, ultrasound, or X-rays [36]. This allows for even greater control over the location and timing of the PS activation, potentially reducing side effects and improving treatment efficacy. Fourth-generation PSs are still a relatively new area of research but have shown promising results in preclinical studies [37–40].

3.3. Natural and Synthetic PS

Herbal medicines have been used for centuries to treat various human ailments, and many modern medicines are derived from medicinal plants. There is a growing interest in using natural products, including herbal extracts and compounds, as possible lead components in cancer drug discoveries. Natural products are a valuable source for the development of drugs and the discovery of new PSs [41–44]. In recent decades, efforts have been made to isolate new products of natural origin from microbes, plants, and other living organisms, leading to the discovery of a number of anti-cancer drugs [41]. Researchers have found new active chemical compounds in natural extracts that display efficient PS properties, and numerous compounds have already been identified as potential candidates to be used as PSs [44]. Furthermore, screening studies of natural extracts have identified new PSs with cyclic tetrapyrrole structures [45]. Additionally, in a 2022 review published by Kubrak TP et al., various photochemicals were described for their light-absorbing properties and their anticancer activity including furanocoumarins, polyacetylenes, thiophenes, curcumins, alkaloids, and anthraquinones [20].

On the other hand, synthetic PSs are those that are created in the laboratory through chemical synthesis. These PSs are designed to have specific properties that allow them to effectively target cancer cells and destroy them upon exposure to light. They are often optimized to have higher light absorption at longer wavelengths, improved tissue penetration, and reduced toxicity compared to earlier generations of PSs [37,46]. Synthetic PSs can be produced in large quantities and with a high degree of purity, making them a reliable and cost-effective option for PDT. Some examples of synthetic PSs include benzoporphyrins, texaphyrins, purpurin, chlorin e6, bacteriochlorin. and pyropheophorbide.

Figure 2 presents some natural and synthetic PSs according to their absorption wavelengths.



Figure 2. Absorption wavelengths of non-tetrapyrrolic PS derivatives (furanocoumarins (Fu), thiophenes (Th), curcumins (Cu), flavonoids (Fl), anthraquinones (An), alkaloids (Al)) and tetrapyrrolic PS derivatives (benzoporphyrin (Be), texaphyrins (Te), purpurin (Pu), chlorin e6 (C6), bacteriochlorin (Ba), pyropheophorbide (Py)).

An updated summary of natural and synthetic PSs and their potential mechanism of action to target prostate cancer are presented in Table 1 and Figure 3.

Table 1. Natural and Synthetic PSs and their potential mechanism of action in PDT of prostate cancer (adapted from Muniyandi K et al. [43]).

Natural PSs					
Name	Natural Sources	Type of PDT *	Mechanism of Action		
Furanocoumarins (Fu)	Ruta graveolens, Angelicae dahuricae, Glehnia littoralis, Syzygium Sps Tetradium daniellii, and Ficus sps.	Type II	 Inhibition of STAT-3 pathway and PI3K/AKT pathway [47,48], Activation of p53 [49], Forming adducts with DNA and cell cycle arrest [50], Upregulation of the mitochondrial genes BAK, BAX, and PUMA [49,51]. 		
Thiophenes (Th)	Echinops, Eclipta, Pluchea, Artemisia, Atractylodes, Tagetes, Porophyllum, and Xanthium.	Type I and II	 Accumulation in fatty acid-rich regions and by producing single oxygen leading to cell membrane destruction [52]. 		
Curcumins (Cu)	Curcuma longa (family Zingiberaceae, species Curcuma)	Type I	 Reduction of matrix metalloproteases' (MMPs) enzyme production [53], Downregulation of inflammatory pathways molecules (e.g., tumor necrosis factor (TNF)-α, lipoxygenase (LOX), and cyclooxygenase (COX)-2) transcription factors (e.g., AP-1, nuclear factor (NF)-κB) and growth factors (e.g., epidermal growth factor (EGFR) and HER-2) [54,55], Activation of caspase-3, -8, and -9 and cytochrome c release [56], Angiogenesis inhibition [44,54,55,57]. 		
Flavonoids (Fl)	Herbs, Vegetables, Seeds, Cereals, Fruits, Flowers, and Nuts	Type II	 ROS generation, DNA damage [58], Upregulate the expression of apoptosis-related proteins [58,59], Increase the expression and release of cytochrome c [59]. 		
Anthraquinones (An)	Polygonum cuspidatum, H. lycioides, Aloe vera, Rheum palmatu, Polyathia suberosa, Xanthoria parietina, Ramularia collo-cygni. and H. perforatum	Type I and II	 ROS generation [60], Induction of apoptosis, necrosis, or autophagy [60,61]. 		
Alkaloids (Al)	Guatteria blepharophylla, Berberis vulgaris, Sanguinaria Canadensis, Peganum harmala, and Indigofera tinctoria.	Type I	 ROS generation [62], Promotion of apoptosis and autophagy by regulating several genes (Bax, Bcl-2, Bcl-xL, NF-κB) and various caspase proteins [62–64]. 		

Synthetic PSs					
Name	Commercial Name/Excitation Wavelength	PS Class/Source	Type of PDT *	Mechanism of Action	
Benzoporphyrin (Be)	Visudyne λ = 689 \pm 3 nm	Porphyrin-based PS (Derived from benzoporphyrin)	Type II	 Lymphocyte infiltration and inhibition of tumor metastasis [26,65], Once excited, it enhances the formation of ·OH from intracellular H₂O₂ via a Fenton-like reaction by a catalyzer [26]. 	
Texaphyrins (Te)	Lutrin, Antrin, Optrin, Xcytrin λ = 732 nm	Texaphyrin-based PS (Derived from expanded porphyrins)	Type I	- Weakens the cellular antioxidant system by inhibiting thioredoxin reductase and by oxidizing reducing species (e.g., ascorbate), leading to a buildup of damaging ROS (e.g., peroxide) [46,66].	
Purpurin (Pu)	Purlytin $\lambda = 664 \text{ nm}$	Porphyrin-based PS (Derived from chlorophyll)	Type II	- Production of singlet oxygen and other ROS that can cause oxidative damage to cellular structures such as lipids, proteins, and DNA and induce apoptosis [67,68].	
Chlorin e6 (C6)	MACEDACEPhotoditazine $\lambda = 660 \text{ nm}$	Chlorin-based PS (Derived from chlorophyll)	Type I and II	- Induce oxidative endoplasmic reticulum stress and DNA damage [69].	
Bacteriochlorin (Ba)	TOOKAD λ = 753 or 762 nm	Chlorin-based PS (Derived from bacteriochlorophyll)	Type I and II	 ROS generation. Blocks blood and nutriment supply to the vascular damage and tumor accumulation [70]. 	
Pyropheophorbide (Py)	Pyropheophorbide-a methylester $\lambda = 675 \text{ nm}$	Chlorin-based PS (Derived from chlorophyll)	Type II	 Targeting prostate-specific membrane antigen (PSMA) [71], ROS generation and apoptosis induction via cytochrome c release, caspase 3 and caspase 9 activation [72]. 	

Table 1. Cont.

* Depending on the solvent used for the measurement [73].

3.4. Ongoing Clinical Studies in Prostate Cancer

In clinical settings, the implementation of PDT for prostate cancer involves several key steps. First, a photosensitizing agent is administered to the patient, either systemically or directly into the tumor site. The choice of photosensitizer depends on factors such as its absorption and distribution properties, as well as its ability to selectively accumulate in tumor tissues [74]. Once the photosensitizer has been adequately distributed, a specific light source is selected, considering factors, such as wavelength and power output [74]. Light delivery techniques may vary, including intraluminal or interstitial illumination [75]. For instance, intraluminal light sources can be used for prostate cancer by inserting light-emitting devices through catheters or endoscopes. Interstitial PDT involves the placement of optical fibers or diffusers directly into the prostate tissue, allowing for light delivery to deeper tissue layers [75]. These techniques aim to achieve optimal light distribution and tumor coverage.

PDT is a promising treatment option for prostate cancer, with the potential to provide a safe and effective alternative to conventional therapies. However, in order to fully evaluate the safety and efficacy of PDT for prostate cancer, clinical trials are needed. These trials can provide valuable insights into the effectiveness of PDT, as well as help to identify any potential side effects or limitations of the treatment.



Figure 3. Possible mechanisms of action to induce prostate cancer eradication by targeting several pathways involving oxidative stress, cell death and cell cycle arrest.

To date, several clinical trials have been conducted on the use of PDT for prostate cancer, with promising results.

By conducting research on clinical trials to further explore the potential of PDT for the treatment of prostate cancer [76], we found that out of the 14 studies conducted (Table 2), using the keywords "Photodynamic therapy" and "Prostate cancer", only one study posted their results [77].

3.5. Multimodal Synergistic Therapies to Overcome PDT Limitations

Combination approaches in PDT for cancer treatment address limitations related to tumor size and location [78]. Larger tumors and tumors located in challenging anatomical areas can pose difficulties in achieving complete eradication with PDT alone due to the limited light penetration depth [79]. However, by combining PDT with other treatment modalities, such as surgery, sonodynamic therapy, photoimmunotherapy, chemotherapy, and photothermal therapy, these limitations can be overcome [80]. For larger tumors, combination approaches can sensitize the tumor or target residual cancer cells, thus enhancing the therapeutic effect. Multimodal therapies that combine PDT with other treatments have shown promise in the treatment of multidrug-resistant and hypoxia-related prostate cancers (Figure 4) [4,81]. However, challenges arise in terms of complexity, potential side effects and the requirement for rigorous testing of optimal sequencing and timing strategies.

Intervention/ Treatment	Phase	Actual Enrollment	Identifier	Responsible Party	Results
Drug: 1 -mediated -VTP WST 1	Phase 1 Phase 2 Terminated	30	NCT00946881	UCLA—Jonsson Comprehensive Cancer Center Los Angeles, California, United States Midtown Urology & Midtown Urology Surgical Center Atlanta, Georgia, United States Washington University School of Medicine- Barnes-Jewish Hospital Saint Louis, Missouri, United States NYU Urology Associates, New York, New York, United States Memorial Sloan-Kettering Cancer Center, New York, New York, United States	The study showed that hemi-ablation of the prostate with WST11 vascular targeted PDT could be an effective treatment for prostate cancer. Optimal dosing parameters and light dose index were found to play important roles in the tissue response, as determined by MRI and biopsy [77].
Drug: WST09	Phase 2 Completed	28	NCT00308919	Princess Margaret Hospital Toronto, Ontario, Canada	No Results Posted
Drug: WST09 Vascular Photodynamic therapy	Phase 2 Phase 3 Terminated	16	NCT00312442	Abramson Cancer Center of The University of Pennsylvania Philadelphia, Pennsylvania, United States	No Results Posted
Drug: WST11	Phase 2 Completed	86	NCT00975429	Centre Hospitalier Universitaire (CHU) Angers, France Hôpital Claude Huriez Lille, France Institut Mutualiste Montsouris (IMM) Paris, France University College London Hospital (UCLH) Kings College Hospital (KCH) Frimley Park Hospital NHS Trust Catharina Ziekenhuis	No Results Posted
Drug: Motexafin lutetium	Phase 1 Terminated	24	NCT00005067	Abramson Cancer Center of The University of Pennsylvania Philadelphia, Pennsylvania, United States	No Results Posted
Drug: TOOKAD [®] Soluble 4 mg/kg	Phase 2 Active	50	NCT03315754	Memorial Sloan-Kettering Cancer Center New York, New York, United States	No Results Posted
Focal therapies including PDT (PS undetermined)	Recruiting	200	NCT03492424	Weill Cornell Medicine New York, New York, United States	No Results Posted
Drug: WST11	Phase 2 Completed	42	NCT00707356	University Health Network-Princess Margaret Hospital Toronto, Canada Centre Hospitalier Universitaire Angers, France Hôpital Bocage-CHU Dijon, France Hôpital Claude Huriez, Lille, France Institut Mutualiste Montsouris (IMM), Paris, France Frimley Park Hospital NHS Trust, Frimley, United Kingdom	No Results Posted
Drug: TOOKAD [®] Soluble	Phase 2 Completed	8	NCT00305929	The Prostate Centre Princess Margaret Hospital Toronto, Ontario, Canada	No Results Posted
Drug: TOOKAD [®] Soluble	Phase 3 Completed	81	NCT01875393	Hospital General Tlahuac Mexico DF, Mexico Pan-American Medical Research Institute (PAMRI) then moved to Consultario del Dr Rodriguez Panama city, Panama Hospital Nacional Cayetano Heredia San Martin de Porres, Peru	No Results Posted

Table 2. An update of clinical studies conducted around the world to treat prostate cancer using PDT and combination approaches.

Intervention/ Treatment	Phase	Actual Enrollment	Identifier	Responsible Party	Results
Drug: TOOKAD [®] Soluble	Phase 3 Withdrawn	0	NCT04225299	Memorial Sloan-Kettering Cancer Center New York, New York, United States	No Results Posted
Drug: TOOKAD [®] Soluble	Phase 4 Terminated	23	NCT03849365	Centre Hospitalier Universitaire Angers, France	No Results Posted
Drug: Visudyne [®] Device: SpectraCure P18 System	Phase 1 Phase 2 Recruiting	66	NCT03067051	Memorial Sloan Kettering Cancer Center New York, New York, United States Keith Cengel Philadelphia, Pennsylvania, United States Princess Margaret Cancer Centre Toronto, Ontario, Canada	No Results Posted
Drug: TOOKAD [®] Soluble	Phase 3 Completed	413	NCT01310894	Dept. of Urology-University Hospitals Leuven Leuven, Belgium Department of Urology-Tampere University Hospital- Tampere, Finland Service d'Urologie—Centre Hospitalier Universitaire Angers, France	No Results Posted

Table 2. Cont.

3.5.1. PDT/Surgery

Surgery is a common treatment for prostate cancer, but it is not always successful in eradicating all cancer cells. Recent studies have explored the potential benefits of combining surgery with PDT in the treatment of prostate cancer. A retrospective study published in 2019 by Pierrard V et al. [82] investigated the safety and effectiveness of salvage radical prostatectomy (surgical removal of the prostate gland) after vascular-targeted photodynamic therapy (VTP) treatment. The study included 45 patients who underwent a salvage prostatectomy followed by VTP salvage due to persistent or recurrent cancer. The results showed that the combination was a safe and effective treatment option for localized prostate cancer. The study also reported that the combination therapy did not result in any severe complications or adverse events [82]. Furthermore, the feasibility, safety, and effectiveness of this combination in treating most locally recurrent prostate cancer were also reported by a previous study where various outcomes were reported including operation time, blood loss, complications, urethral catheterization time, functional outcomes, and short-term oncologic outcomes [83].

3.5.2. PDT/Sonodynamic Therapy

Although there are no specific studies on the use of PDT and sonodynamic therapy (SDT) for prostate cancer treatment, these therapies are currently being studied for their potential in cancer treatments [39,84]. PDT and SDT utilize sensitizers that can produce ROS, which can cause damage to cancer cell membranes. The combination of these therapies has demonstrated promising results in preclinical studies for various types of cancer, such as brain cancer [39]. In fact, in the prostate cancer context, the penetrating power of ultrasound avoids the use of an endoscope (and light) in the bladder while allowing the activation of the PS [84]. Ongoing research aims to identify effective sensitizers and optimal delivery methods to enhance the effectiveness of PDT and SDT in prostate cancer treatments. While further studies are necessary to determine the safety and efficacy of these therapies for prostate cancer, the potential benefits of PDT and SDT offer a promising avenue for the development of effective cancer treatments.

3.5.3. PDT/Photoimmunotherapy

Photoimmunotherapy (PIT) is an innovative treatment that harnesses the power of phototherapy and immunotherapy to combat cancer by inducing antitumor immune responses and preventing cancer recurrence [85]. Among the different approaches to PIT, one of the most promising and rapidly expanding is near-infrared (NIR)-PIT. This approach selectively and locally destroys cancer cells by releasing damage-associated molecular patterns (DAMPs) and tumor-associated antigens (TAAs), which trigger an antitumor immune response. NIR-PIT has demonstrated remarkable results in preclinical and clinical applications for various types of cancer including prostate cancer [85,86]. In fact, in a study using a prostate-specific membrane antigen (PSMA)-expressing a prostate cancer cell line, NIR-PIT with a fully human IgG1 anti-PSMA monoclonal antibody (mAb) conjugated to a specific photoabsorber resulted in specific binding and cell-specific killing after exposure to NIR light [86].

3.5.4. PDT/Chemotherapy

In the last few years, researchers have developed a new treatment strategy for prostate cancer known as photochemotherapy. This approach combines the benefits of PDT and chemotherapy to deliver cancer-fighting drugs selectively to tumor cells. Unlike traditional chemotherapy, photochemotherapy can target cancerous tissues without damaging surrounding healthy cells, potentially reducing unwanted side effects. By combining these two therapies, the effectiveness of the treatment is enhanced, with chemotherapy may increase the sensitivity of cancer cells to ROS or hyperthermia, resulting in a synergistic effect [87]. Furthermore, in vivo studies using athymic nude mice and BALB/c nude mice, have also demonstrated the effectiveness of photochemotherapy in treating prostate cancer [88,89].

3.5.5. PDT/Photothermal Therapy

Photothermal therapy (PTT) is a form of phototherapy that uses light energy to produce heat energy, which can kill cancer cells [4]. PTT has been used to treat prostate cancer by irradiating a photothermal agent with an external light source of a specific wavelength, typically NIR light. PTT has been combined with PDT to enhance the effects of both therapies. The thermal effect of PTT can improve local blood flow and increase the oxygen concentration in the tumor tissue, making up for the deficiency of PDT in a hypoxic environment [90]. Many studies have reported the combination of PDT and PTT using nanomaterials as drug carriers [4,90]. Some studies have used self-assembled nanoparticles, such as melanin-like polydopamine nanoparticles, to deliver photothermal and photodynamic agents for synergistic therapy [4,90]. However, the low photoconversion efficiency of some materials used in these therapies raises issues about how to maximize the synergistic effect of PDT and PTT.



Figure 4. Schematic illustration of mechanisms of the multimodal synergistic therapies targeting prostate cancer using PDT.

4. Conclusions

In the last decades, steady progress in prostate cancer patients' care has been noted due to the broad application of early detection and progress in therapeutic approaches. In this context, PDT is a promising approach to treat prostate cancer that has the potential to provide a minimally invasive, highly targeted approach and negligible side effects on normal tissues when compared to conventional therapies. Several PDT approaches have already been approved by regulatory authorities including the FDA and the European Medicines Agency. Furthermore, combining PDT with other therapy modalities such as chemotherapy, radiotherapy, surgery, and immunotherapy seems to be more effective against tumor growth. Advances in therapeutic approaches to develop innovative organic PSs for enhanced PDT are being reported worldwide [91]. Recently, PDT has entered the era of vectorization and encapsulation into drug delivery systems using innovative strategies to increase the delivery of PSs to tumor tissues [89,92–95]. Furthermore, several research studies reported the use of PDT to treat different cancer types, including colorectal cancer, melanoma, and glioblastoma as well as other diseases such as rheumatoid arthritis [92–97]. Despite considerable progress in understanding the underlying mechanisms of photodynamic therapy, further clinical studies are still needed to improve its modalities.

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References

- Sung, H.; Ferlay, J.; Siegel, R.L.; Laversanne, M.; Soerjomataram, I.; Jemal, A.; Bray, F. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. CA Cancer J. Clin. 2021, 71, 209–249. [CrossRef]
- Algorri, J.F.; Ochoa, M.; Roldan-Varona, P.; Rodriguez-Cobo, L.; Lopez-Higuera, J.M. Photodynamic Therapy: A Compendium of Latest Reviews. *Cancers* 2021, 13, 4447. [CrossRef] [PubMed]
- Windahl, T.; Andersson, S.O.; Lofgren, L. Photodynamic therapy of localised prostatic cancer. *Lancet* 1990, 336, 1139. [CrossRef] [PubMed]
- Xue, Q.; Zhang, J.; Jiao, J.; Qin, W.; Yang, X. Photodynamic therapy for prostate cancer: Recent advances, challenges and opportunities. *Front. Oncol.* 2022, 12, 980239. [CrossRef]
- 5. PDQ[®] Adult Treatment Editorial Board. Prostate Cancer Treatment: Health Professional Version. Updated: 13 February 2023. Available online: https://www.cancer.gov/types/prostate/hp/prostate-treatment-pdq (accessed on 14 May 2023).
- 6. Mohan, R.; Schellhammer, P.F. Treatment options for localized prostate cancer. Am. Fam. Physician 2011, 84, 413–420.
- 7. Sekhoacha, M.; Riet, K.; Motloung, P.; Gumenku, L.; Adegoke, A.; Mashele, S. Prostate Cancer Review: Genetics, Diagnosis, Treatment Options, and Alternative Approaches. *Molecules* **2022**, *27*, 5730. [CrossRef]
- 8. Wallis, C.J.; Herschorn, S.; Saskin, R.; Su, J.; Klotz, L.H.; Chang, M.; Kulkarni, G.S.; Lee, Y.; Kodama, R.T.; Narod, S.A.; et al. Complications after radical prostatectomy or radiotherapy for prostate cancer: Results of a population-based, propensity score-matched analysis. *Urology* **2015**, *85*, 621–627. [CrossRef]
- 9. Michaelson, M.D.; Cotter, S.E.; Gargollo, P.C.; Zietman, A.L.; Dahl, D.M.; Smith, M.R. Management of Complications of Prostate Cancer Treatment. *CA Cancer J. Clin.* 2008, *58*, 196–213. [CrossRef]
- 10. Garcia, J.A. Sipuleucel-T in patients with metastatic castration-resistant prostate cancer: An insight for oncologists. *Ther. Adv. Med. Oncol.* **2011**, *3*, 101–108. [CrossRef]
- 11. Fitzgerald, F. Photodynamic Therapy (PDT); Nova Science Publishers, Incorporated: Hauppauge, NY, USA, 2017.
- 12. Correia, J.H.; Rodrigues, J.A.; Pimenta, S.; Dong, T.; Yang, Z. Photodynamic Therapy Review: Principles, Photosensitizers, Applications, and Future Directions. *Pharmaceutics* **2021**, *13*, 1332. [CrossRef] [PubMed]
- 13. Mishchenko, T.; Balalaeva, I.; Gorokhova, A.; Vedunova, M.; Krysko, D.V. Which cell death modality wins the contest for photodynamic therapy of cancer? *Cell Death Dis.* **2022**, *13*, 455. [CrossRef]
- 14. Hamblin, M.R. Photodynamic Therapy for Cancer: What's Past is Prologue. *Photochem. Photobiol.* **2020**, *96*, 506–516. [CrossRef] [PubMed]
- 15. Lee, C.N.; Hsu, R.; Chen, H.; Wong, T.W. Daylight Photodynamic Therapy: An Update. Molecules 2020, 25, 5195. [CrossRef]
- Rodrigues, M.C.; de Sousa Junior, W.T.; Mundim, T.; Vale, C.L.C.; de Oliveira, J.V.; Ganassin, R.; Pacheco, T.J.A.; Vasconcelos Morais, J.A.; Longo, J.P.F.; Azevedo, R.B.; et al. Induction of Immunogenic Cell Death by Photodynamic Therapy Mediated by Aluminum-Phthalocyanine in Nanoemulsion. *Pharmaceutics* 2022, 14, 196. [CrossRef] [PubMed]
- 17. Rocha, L.G.B. Development of a Novel Photosensitizer for Photodynamic Therapy of Cancer. Ph.D. Thesis, Universidade de Coimbra, Coimbra, Portugal, 2016.
- Kwiatkowski, S.; Knap, B.; Przystupski, D.; Saczko, J.; Kedzierska, E.; Knap-Czop, K.; Kotlinska, J.; Michel, O.; Kotowski, K.; Kulbacka, J. Photodynamic therapy—Mechanisms, photosensitizers and combinations. *Biomed. Pharmacother.* 2018, 106, 1098–1107. [CrossRef]
- 19. Mroz, P.; Yaroslavsky, A.; Kharkwal, G.B.; Hamblin, M.R. Cell death pathways in photodynamic therapy of cancer. *Cancers* **2011**, *3*, 2516–2539. [CrossRef]
- Kubrak, T.P.; Kołodziej, P.; Sawicki, J.; Mazur, A.; Koziorowska, K.; Aebisher, D. Some Natural Photosensitizers and Their Medicinal Properties for Use in Photodynamic Therapy. *Molecules* 2022, 27, 1192. [CrossRef] [PubMed]
- Zhang, J.; Jiang, C.; Figueiró Longo, J.P.; Azevedo, R.B.; Zhang, H.; Muehlmann, L.A. An updated overview on the development of new photosensitizers for anticancer photodynamic therapy. *Acta Pharm. Sin. B* 2018, *8*, 137–146. [CrossRef]
- 22. Oleinick, N.L.; Morris, R.L.; Belichenko, I. The role of apoptosis in response to photodynamic therapy: What, where, why, and how. *Photochem. Photobiol. Sci.* 2002, 1, 1–21. [CrossRef]

- Tomasz, K.; Beata, C.-G.; Katarzyna, K.; Magdalena, S.; Daria, W.; Sebastian, L.; Tomasz, K.; Tomasz, G. Nurses and Pharmacists in Interdisciplinary Team of Health Care Providers in Photodynamic Therapy. In *Photomedicine*; Yohei, T., Ed.; IntechOpen: Rijeka, Croatia, 2017; Chapter 7. [CrossRef]
- 24. O'Connor, A.E.; Gallagher, W.M.; Byrne, A.T. Porphyrin and nonporphyrin photosensitizers in oncology: Preclinical and clinical advances in photodynamic therapy. *Photochem. Photobiol.* **2009**, *85*, 1053–1074. [CrossRef] [PubMed]
- 25. Nayak, C.S. Photodynamic therapy in dermatology. Indian J. Dermatol. Venereol. Leprol. 2005, 71, 155–160. [CrossRef] [PubMed]
- Zeng, J.Y.; Zou, M.Z.; Zhang, M.; Wang, X.S.; Zeng, X.; Cong, H.; Zhang, X.Z. π-Extended Benzoporphyrin-Based Metal—Organic Framework for Inhibition of Tumor Metastasis. ACS Nano 2018, 12, 4630–4640. [CrossRef]
- 27. Dai, L.; Shen, G.; Wang, Y.; Yang, P.; Wang, H.; Liu, Z. PSMA-targeted melanin-like nanoparticles as a multifunctional nanoplatform for prostate cancer theranostics. *J. Mater. Chem. B* **2021**, *9*, 1151–1161. [CrossRef]
- Stallivieri, A.; Colombeau, L.; Jetpisbayeva, G.; Moussaron, A.; Myrzakhmetov, B.; Arnoux, P.; Acherar, S.; Vanderesse, R.; Frochot, C. Folic acid conjugates with photosensitizers for cancer targeting in photodynamic therapy: Synthesis and photophysical properties. *Bioorganic Med. Chem.* 2017, 25, 1–10. [CrossRef]
- Stallivieri, A.; Baros, F.; Jetpisbayeva, G.; Myrzakhmetov, B.; Frochot, C. The Interest of Folic Acid in Targeted Photodynamic Therapy. Curr. Med. Chem. 2015, 22, 3185–3207. [CrossRef]
- Singh, S.; Aggarwal, A.; Bhupathiraju, N.V.S.D.K.; Arianna, G.; Tiwari, K.; Drain, C.M. Glycosylated Porphyrins, Phthalocyanines, and Other Porphyrinoids for Diagnostics and Therapeutics. *Chem. Rev.* 2015, 115, 10261–10306. [CrossRef]
- Laville, I.; Figueiredo, T.; Loock, B.; Pigaglio, S.; Maillard, P.; Grierson, D.S.; Carrez, D.; Croisy, A.; Blais, J. Synthesis, cellular internalization and photodynamic activity of glucoconjugated derivatives of tri and tetra(meta-hydroxyphenyl)chlorins. *Bioorganic Med. Chem.* 2003, *11*, 1643–1652. [CrossRef] [PubMed]
- Kamarulzaman, E.E.; Gazzali, A.M.; Acherar, S.; Frochot, C.; Barberi-Heyob, M.; Boura, C.; Chaimbault, P.; Sibille, E.; Wahab, H.A.; Vanderesse, R. New Peptide-Conjugated Chlorin-Type Photosensitizer Targeting Neuropilin-1 for Anti-Vascular Targeted Photodynamic Therapy. *Int. J. Mol. Sci.* 2015, *16*, 24059–24080. [CrossRef]
- 33. Chaleix, V.; Sol, V.; Huang, Y.-M.; Guilloton, M.; Granet, R.; Blais, J.C.; Krausz, P. RGD-Porphyrin Conjugates: Synthesis and Potential Application in Photodynamic Therapy. *Eur. J. Org. Chem.* **2003**, 2003, 1486–1493. [CrossRef]
- 34. Sol, V.; Lamarche, F.; Enache, M.; Garcia, G.; Granet, R.; Guilloton, M.; Blais, J.C.; Krausz, P. Polyamine conjugates of mesotritolylporphyrin and protoporphyrin IX: Potential agents for photodynamic therapy of cancers. *Bioorganic Med. Chem.* **2006**, *14*, 1364–1377. [CrossRef]
- 35. Garcia, G.; Sarrazy, V.; Sol, V.; Morvan, C.L.; Granet, R.; Alves, S.; Krausz, P. DNA photocleavage by porphyrin–polyamine conjugates. *Bioorganic Med. Chem.* 2009, 17, 767–776. [CrossRef]
- Bechet, D.; Couleaud, P.; Frochot, C.; Viriot, M.L.; Guillemin, F.; Barberi-Heyob, M. Nanoparticles as vehicles for delivery of photodynamic therapy agents. *Trends Biotechnol.* 2008, 26, 612–621. [CrossRef]
- 37. Lismont, M.; Dreesen, L.; Wuttke, S. Metal-Organic Framework Nanoparticles in Photodynamic Therapy: Current Status and Perspectives. *Adv. Funct. Mater.* **2017**, *27*, 1606314. [CrossRef]
- Foglietta, F.; Pinnelli, V.; Giuntini, F.; Barbero, N.; Panzanelli, P.; Durando, G.; Terreno, E.; Serpe, L.; Canaparo, R. Sonodynamic Treatment Induces Selective Killing of Cancer Cells in an In Vitro Co-Culture Model. *Cancers* 2021, 13, 3852. [CrossRef]
- Borah, B.M.; Cacaccio, J.; Durrani, F.A.; Bshara, W.; Turowski, S.G.; Spernyak, J.A.; Pandey, R.K. Sonodynamic therapy in combination with photodynamic therapy shows enhanced long-term cure of brain tumor. *Sci. Rep.* 2020, 10, 21791. [CrossRef]
- Bulin, A.-L.; Truillet, C.; Chouikrat, R.; Lux, F.; Frochot, C.; Amans, D.; Ledoux, G.; Tillement, O.; Perriat, P.; Barberi-Heyob, M.; et al. X-ray-Induced Singlet Oxygen Activation with Nanoscintillator-Coupled Porphyrins. *J. Phys. Chem. C* 2013, 117, 21583–21589. [CrossRef]
- Newman, D.J.; Cragg, G.M. Natural Products as Sources of New Drugs over the Nearly Four Decades from 01/1981 to 09/2019. J. Nat. Prod. 2020, 83, 770–803. [CrossRef]
- 42. Zhang, Y.; Huang, L.; Shi, H.; Chen, H.; Tao, J.; Shen, R.; Wang, T. Ursolic acid enhances the therapeutic effects of oxaliplatin in colorectal cancer by inhibition of drug resistance. *Cancer Sci.* **2018**, *109*, 94–102. [CrossRef]
- Muniyandi, K.; George, B.; Parimelazhagan, T.; Abrahamse, H. Role of Photoactive Phytocompounds in Photodynamic Therapy of Cancer. *Molecules* 2020, 25, 4102. [CrossRef]
- 44. Mansoori, B.; Mohammadi, A.; Doustvandi, M.A.; Mohammadnejad, F.; Kamari, F.; Gjerstorff, M.F.; Baradaran, B.; Hamblin, M.R. Photodynamic therapy for cancer: Role of natural products. *Photodiagnosis Photodyn. Ther.* **2019**, *26*, 395–404. [CrossRef]
- 45. Samat, N.; Tan, P.J.; Shaari, K.; Abas, F.; Lee, H.B. Prioritization of natural extracts by LC-MS-PCA for the identification of new photosensitizers for photodynamic therapy. *Anal. Chem.* **2014**, *86*, 1324–1331. [CrossRef] [PubMed]
- 46. Arambula, J.F.; Preihs, C.; Borthwick, D.; Magda, D.; Sessler, J.L. Texaphyrins: Tumor localizing redox active expanded porphyrins. *Anticancer Agents Med. Chem.* **2011**, *11*, 222–232. [CrossRef]
- Panno, M.L.; Giordano, F.; Rizza, P.; Pellegrino, M.; Zito, D.; Giordano, C.; Mauro, L.; Catalano, S.; Aquila, S.; Sisci, D.; et al. Bergapten induces ER depletion in breast cancer cells through SMAD4-mediated ubiquitination. *Breast Cancer Res. Treat.* 2012, 136, 443–455. [CrossRef]

- Van Aelst, B.; Devloo, R.; Zachee, P.; t'Kindt, R.; Sandra, K.; Vandekerckhove, P.; Compernolle, V.; Feys, H.B. Psoralen and Ultraviolet A Light Treatment Directly Affects Phosphatidylinositol 3-Kinase Signal Transduction by Altering Plasma Membrane Packing. J. Biol. Chem. 2016, 291, 24364–24376. [CrossRef]
- Mastrangelopoulou, M.; Grigalavicius, M.; Berg, K.; Menard, M.; Theodossiou, T.A. Cytotoxic and Photocytotoxic Effects of Cercosporin on Human Tumor Cell Lines. *Photochem. Photobiol.* 2019, 95, 387–396. [CrossRef]
- Ge, Z.-C.; Qu, X.; Yu, H.-F.; Zhang, H.-M.; Wang, Z.-H.; Zhang, Z.-T. Antitumor and apoptotic effects of bergaptol are mediated via mitochondrial death pathway and cell cycle arrest in human breast carcinoma cells. *Bangladesh J. Pharmacol.* 2016, 11, 489–494. [CrossRef]
- Liszewski, W.; Naym, D.G.; Biskup, E.; Gniadecki, R. Psoralen with ultraviolet A-induced apoptosis of cutaneous lymphoma cell lines is augmented by type I interferons via the JAK1-STAT1 pathway. *Photodermatol. Photoimmunol. Photomed.* 2017, 33, 164–171. [CrossRef] [PubMed]
- 52. Simpson, D.; Amos, S. Other plant metabolites. In *Pharmacognosy*; Elsevier: Cham, Switzerland, 2017; pp. 267–280.
- 53. Priyadarsini, K.I. Photophysics, photochemistry and photobiology of curcumin: Studies from organic solutions, bio-mimetics and living cells. J. Photochem. Photobiol. C Photochem. Rev. 2009, 10, 81–95. [CrossRef]
- 54. Abrahamse, H.; Hamblin, M.R. New photosensitizers for photodynamic therapy. Biochem. J. 2016, 473, 347–364. [CrossRef]
- 55. Aziz, B.; Aziz, I.; Khurshid, A.; Raoufi, E.; Esfahani, F.N.; Jalilian, Z.; Mozafari, M.R.; Taghavi, E.; Ikram, M. An Overview of Potential Natural Photosensitizers in Cancer Photodynamic Therapy. *Biomedicines* **2023**, *11*, 224. [CrossRef]
- 56. Park, K.; Lee, J.-H. Photosensitizer effect of curcumin on UVB-irradiated HaCaT cells through activation of caspase pathways. *Oncol. Rep.* **2007**, *17*, 537–540. [CrossRef]
- 57. Mbese, Z.; Khwaza, V.; Aderibigbe, B.A. Curcumin and Its Derivatives as Potential Therapeutic Agents in Prostate, Colon and Breast Cancers. *Molecules* **2019**, *24*, 4386. [CrossRef]
- 58. Zhang, S.-J.; Sun, D.; Hao, J.-B.; Wei, Y.-F.; Yin, L.-F.; Liu, X. The effect of dietary soyabean isoflavones on photodynamic therapy in K562 leukemia cells. *J. Photochem. Photobiol. B Biol.* **2012**, *110*, 28–33. [CrossRef] [PubMed]
- 59. de Paula Rodrigues, R.; Tini, I.R.; Soares, C.P.; da Silva, N.S. Effect of photodynamic therapy supplemented with quercetin in HEp-2 cells. *Cell Biol. Int.* **2014**, *38*, 716–722. [CrossRef]
- Núñez Montoya, S.C.; Comini, L.R.; Rumie Vittar, B.; Fernández, I.M.; Rivarola, V.A.; Cabrera, J.L. Phototoxic effects of Heterophyllaea pustulata (Rubiaceae). *Toxicon* 2008, 51, 1409–1415. [CrossRef] [PubMed]
- Vittar, N.B.R.; Awruch, J.; Azizuddin, K.; Rivarola, V. Caspase-independent apoptosis, in human MCF-7c3 breast cancer cells, following photodynamic therapy, with a novel water-soluble phthalocyanine. *Int. J. Biochem. Cell Biol.* 2010, 42, 1123–1131. [CrossRef] [PubMed]
- 62. Perez Martin, J.M.; Labrador, V.; Fernandez Freire, P.; Molero, M.L.; Hazen, M.J. Ultrastructural changes induced in HeLa cells after phototoxic treatment with harmine. *J. Appl. Toxicol.* **2004**, *24*, 197–201. [CrossRef]
- 63. Burgeiro, A.; Gajate, C.; Dakir, E.L.H.; Villa-Pulgarín, J.A.; Oliveira, P.J.; Mollinedo, F. Involvement of mitochondrial and B-RAF/ERK signaling pathways in berberine-induced apoptosis in human melanoma cells. *Anti-Cancer Drugs* **2011**, 22, 507–518. [CrossRef]
- 64. Youn, M.-J.; So, H.-S.; Cho, H.-J.; Kim, H.-J.; Kim, Y.; Lee, J.-H.; Sohn, J.S.; Kim, Y.K.; Chung, S.-Y.; Park, R. Berberine, a Natural Product, Combined with Cisplatin Enhanced Apoptosis through a Mitochondria/Caspase-Mediated Pathway in HeLa Cells. *Biol. Pharm. Bull.* **2008**, *31*, 789–795. [CrossRef]
- 65. Lan, G.; Ni, K.; Xu, Z.; Veroneau, S.S.; Song, Y.; Lin, W. Nanoscale Metal-Organic Framework Overcomes Hypoxia for Photodynamic Therapy Primed Cancer Immunotherapy. J. Am. Chem. Soc. 2018, 140, 5670–5673. [CrossRef]
- 66. Wondrak, G.T. Redox-directed cancer therapeutics: Molecular mechanisms and opportunities. *Antioxid. Redox Signal.* **2009**, *11*, 3013–3069. [CrossRef]
- Pavlickova, V.; Rimpelova, S.; Jurasek, M.; Zaruba, K.; Fahnrich, J.; Krizova, I.; Bejcek, J.; Rottnerova, Z.; Spiwok, V.; Drasar, P.; et al. PEGylated Purpurin 18 with Improved Solubility: Potent Compounds for Photodynamic Therapy of Cancer. *Molecules* 2019, 24, 4477. [CrossRef]
- 68. Huang, P.; Zhang, B.; Yuan, Q.; Zhang, X.; Leung, W.; Xu, C. Photodynamic treatment with purpurin 18 effectively inhibits triple negative breast cancer by inducing cell apoptosis. *Lasers Med. Sci.* **2021**, *36*, 339–347. [CrossRef]
- 69. Yu, T.T.; Hu, J.; Li, Q.R.; Peng, X.C.; Xu, H.Z.; Han, N.; Li, L.G.; Yang, X.X.; Xu, X.; Yang, Z.Y.; et al. Chlorin e6-induced photodynamic effect facilitates immunogenic cell death of lung cancer as a result of oxidative endoplasmic reticulum stress and DNA damage. *Int. Immunopharmacol.* **2023**, *115*, 109661. [CrossRef]
- 70. Azzouzi, A.R.; Lebdai, S.; Benzaghou, F.; Stief, C. Vascular-targeted photodynamic therapy with TOOKAD®Soluble in localized prostate cancer: Standardization of the procedure. *World J. Urol.* **2015**, *33*, 937–944. [CrossRef]
- Harmatys, K.M.; Overchuk, M.; Chen, J.; Ding, L.; Chen, Y.; Pomper, M.G.; Zheng, G. Tuning Pharmacokinetics to Improve Tumor Accumulation of a Prostate-Specific Membrane Antigen-Targeted Phototheranostic Agent. *Bioconjugate Chem.* 2018, 29, 3746–3756. [CrossRef]
- 72. Matroule, J.-Y.; Carthy, C.M.; Granville, D.J.; Jolois, O.; Hunt, D.W.C.; Piette, J. Mechanism of colon cancer cell apoptosis mediated by pyropheophorbide-a methylester photosensitization. *Oncogene* **2001**, *20*, 4070–4084. [CrossRef]

- 73. Nyman, E.S.; Hynninen, P.H. Research advances in the use of tetrapyrrolic photosensitizers for photodynamic therapy. *J. Photochem. Photobiol. B* **2004**, *73*, 1–28. [CrossRef]
- van Straten, D.; Mashayekhi, V.; de Bruijn, H.S.; Oliveira, S.; Robinson, D.J. Oncologic Photodynamic Therapy: Basic Principles, Current Clinical Status and Future Directions. *Cancers* 2017, 9, 19. [CrossRef] [PubMed]
- Moore, C.M.; Pendse, D.; Emberton, M. Photodynamic therapy for prostate cancer—A review of current status and future promise. *Nat. Clin. Pract. Urol.* 2009, 6, 18–30. [CrossRef]
- 76. Available online: https://clinicaltrials.gov/ (accessed on 14 May 2023).
- 77. Taneja, S.S.; Bennett, J.; Coleman, J.; Grubb, R.; Andriole, G.; Reiter, R.E.; Marks, L.; Azzouzi, A.R.; Emberton, M. Final Results of a Phase I/II Multicenter Trial of WST11 Vascular Targeted Photodynamic Therapy for Hemi-Ablation of the Prostate in Men with Unilateral Low Risk Prostate Cancer Performed in the United States. J. Urol. 2016, 196, 1096–1104. [CrossRef]
- Ito, Y.; Udo, K.; Vertosick, E.A.; Sjoberg, D.D.; Vickers, A.J.; Al-Ahmadie, H.A.; Chen, Y.B.; Gopalan, A.; Sirintrapun, S.J.; Tickoo, S.K.; et al. Clinical Usefulness of Prostate and Tumor Volume Related Parameters following Radical Prostatectomy for Localized Prostate Cancer. J. Urol. 2019, 201, 535–540. [CrossRef]
- 79. Osuchowski, M.; Aebisher, D.; Bartusik-Aebisher, D.; Krupka-Olek, M.; Dynarowicz, K.; Przygoda, M.; Kawczyk-Krupka, A. Photodynamic Therapy-Adjunctive Therapy in the Treatment of Prostate Cancer. *Diagnostics* **2022**, *12*, 1113. [CrossRef]
- 80. Borgia, F.; Giuffrida, R.; Caradonna, E.; Vaccaro, M.; Guarneri, F.; Cannavo, S.P. Early and Late Onset Side Effects of Photodynamic Therapy. *Biomedicines* **2018**, *6*, 12. [CrossRef]
- Sarbadhikary, P.; George, B.P.; Abrahamse, H. Recent Advances in Photosensitizers as Multifunctional Theranostic Agents for Imaging-Guided Photodynamic Therapy of Cancer. *Theranostics* 2021, *11*, 9054–9088. [CrossRef]
- Pierrard, V.; Lebdai, S.; Kleinclauss, F.; Azzouzi, A.-R.; Terrier, J.-E.; Fortier, E.; Joniau, S.; Van Der Poel, H.; Salomon, G.; Casanova, J.; et al. Radical Prostatectomy after Vascular Targeted Photodynamic Therapy with Padeliporfin: Feasibility, and Early and Intermediate Results. *J. Urol.* 2019, 201, 315–321. [CrossRef]
- 83. Lebdai, S.; Villers, A.; Barret, E.; Nedelcu, C.; Bigot, P.; Azzouzi, A.R. Feasibility, safety, and efficacy of salvage radical prostatectomy after Tookad®Soluble focal treatment for localized prostate cancer. *World J. Urol.* **2015**, *33*, 965–971. [CrossRef]
- Hu, X.; Zhang, Y.-S.; Liu, Y.-C.; Wang, N.; Zeng, X.-T.; Zhang, L.-L. Emerging photodynamic/sonodynamic therapies for urological cancers: Progress and challenges. J. Nanobiotechnol. 2022, 20, 437. [CrossRef]
- Mohiuddin, T.M.; Zhang, C.; Sheng, W.; Al-Rawe, M.; Zeppernick, F.; Meinhold-Heerlein, I.; Hussain, A.F. Near Infrared Photoimmunotherapy: A Review of Recent Progress and Their Target Molecules for Cancer Therapy. *Int. J. Mol. Sci.* 2023, 24, 2655. [CrossRef]
- Nagaya, T.; Nakamura, Y.; Okuyama, S.; Ogata, F.; Maruoka, Y.; Choyke, P.L.; Kobayashi, H. Near-Infrared Photoimmunotherapy Targeting Prostate Cancer with Prostate-Specific Membrane Antigen (PSMA) AntibodyNear-Infrared Photoimmunotherapy Targeting PSMA. *Mol. Cancer Res.* 2017, *15*, 1153–1162. [CrossRef]
- Poudel, B.K.; Soe, Z.C.; Ruttala, H.B.; Gupta, B.; Ramasamy, T.; Thapa, R.K.; Gautam, M.; Ou, W.; Nguyen, H.T.; Jeong, J.-H.; et al. In situ fabrication of mesoporous silica-coated silver-gold hollow nanoshell for remotely controllable chemo-photothermal therapy via phase-change molecule as gatekeepers. *Int. J. Pharm.* 2018, 548, 92–103. [CrossRef]
- Wang, X.; Tong, J.; He, Z.; Yang, X.; Meng, F.; Liang, H.; Zhang, X.; Luo, L. Paclitaxel-Potentiated Photodynamic Theranostics for Synergistic Tumor Ablation and Precise Anticancer Efficacy Monitoring. ACS Appl. Mater. Interfaces 2020, 12, 5476–5487. [CrossRef]
- Tan, H.; Hou, N.; Liu, Y.; Liu, B.; Cao, W.; Zheng, D.; Li, W.; Liu, Y.; Xu, B.; Wang, Z.; et al. CD133 antibody targeted delivery of gold nanostars loading IR820 and docetaxel for multimodal imaging and near-infrared photodynamic/photothermal/chemotherapy against castration resistant prostate cancer. *Nanomed. Nanotechnol. Biol. Med.* 2020, 27, 102192. [CrossRef]
- Huang, L.; Xu, C.; Xu, P.; Qin, Y.; Chen, M.; Feng, Q.; Pan, J.; Cheng, Q.; Liang, F.; Wen, X.; et al. Intelligent Photosensitive Mesenchymal Stem Cells and Cell-Derived Microvesicles for Photothermal Therapy of Prostate Cancer. *Nanotheranostics* 2019, 3, 41–53. [CrossRef]
- 91. Pham, T.C.; Nguyen, V.-N.; Choi, Y.; Lee, S.; Yoon, J. Recent Strategies to Develop Innovative Photosensitizers for Enhanced Photodynamic Therapy. *Chem. Rev.* 2021, *121*, 13454–13619. [CrossRef]
- 92. Gallardo-Villagrán, M.; Paulus, L.; Charissoux, J.L.; Sutour, S.; Vergne-Salle, P.; Leger, D.Y.; Liagre, B.; Therrien, B. Evaluation of Ruthenium-Based Assemblies as Carriers of Photosensitizers to Treat Rheumatoid Arthritis by Photodynamic Therapy. *Pharmaceutics* **2021**, *13*, 2104. [CrossRef]
- Nkune, N.W.; Matlou, G.G.; Abrahamse, H. Photodynamic Therapy Efficacy of Novel Zinc Phthalocyanine Tetra Sodium 2-Mercaptoacetate Combined with Cannabidiol on Metastatic Melanoma. *Pharmaceutics* 2022, 14, 2418. [CrossRef]
- Mkhobongo, B.; Chandran, R.; Abrahamse, H. In Vitro Photodynamic Treatment Modality for A375 Melanoma Cell Line Using a Sulphonated Aluminum Phthalocyanine Chloride-Photosensitizer-Gold Nanoparticle Conjugate. *Pharmaceutics* 2022, 14, 2474. [CrossRef]
- Dandash, F.; Leger, D.Y.; Diab-Assaf, M.; Sol, V.; Liagre, B. Porphyrin/Chlorin Derivatives as Promising Molecules for Therapy of Colorectal Cancer. *Molecules* 2021, 26, 7268. [CrossRef]

- 96. Cramer, S.W.; Chen, C.C. Photodynamic Therapy for the Treatment of Glioblastoma. Front. Surg. 2019, 6, 81. [CrossRef]
- 97. Gallardo-Villagrán, M.; Leger, D.Y.; Liagre, B.; Therrien, B. Photosensitizers Used in the Photodynamic Therapy of Rheumatoid Arthritis. *Int. J. Mol. Sci.* 2019, 20, 3339. [CrossRef]

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